

International Journal of Advanced Research in Science, Communication and Technology (IJARSCT)

Volume 3, Issue 3, January 2023

# Significance of Forensic Fly and Seasonal Variation in the Temperature of Developmental Stages in Lifecycle for Family Sarcophagidae, *Sarcophaga Africa*

Dr. Bhosale P.A.

Department of Zoology Sundarrao More Arts, Commerce, and Science (Sr) College, Poladpur, Raigad, Maharashtra, India bhosale popat@rediffmail.com

Abstract: Sarcophaga Africa, is one of the hairy maggot flesh flies which feeds on meat carrion, and dead and decaying matter of animals to the completes its life cycle which is the useful for post mortem interval (PMI). Determination in forensic investigations. The actual life cycle hours and days are calculated due to their morphological parameter of their life cycle of Sarcophaga Africa were studied in different seasons; Life cycle in rainy season was completed in  $270\pm 1.25$  hrs ( $11.25 \pm 0.40$  days), when the maximum temperature was  $27.02^{\circ}$ C and the minimum temperature was  $26.4^{\circ}$ C; in summer season when the maximum temperature was  $36.6^{\circ}$ C and the minimum temperature  $33.2^{\circ}$ C, the life cycle was completed in  $220\pm 1.17$ hrs ( $6.16. \pm 0.10$  days), while in cycle was completed in  $310 \pm 1.35$ hrs ( $12.91 \pm 0.21$  days) when the maximum and minimum winter season life cm temperatures were  $27.4^{\circ}$ C and  $17.2^{\circ}$ C respectively. The temperature is the important role to determine the developmental stages of life cycle of Sarcophaga Africa which should be considered during PMI determination. The external parameters of different stages differ from season to season. Larvae were healthy and bigger in size in rainy season but in summer were short and small sized. The size of larvae in winter season was also smaller than the size in both summer and rainy seasons.

Keywords: Forensic Insect, PMI season, lifecycle duration; Temp change.

### I. INTRODUCTION

Forensic entomology deals with entomological evidence that is relevant in legal cases, particularly those related to corpses. Proper identification of evidence is crucial as a misidentification may lead to inaccurate and erroneous conclusions of potentially dramatic consequences. Identification is usually made on the basis of morphological characters observed on adults and compiled in identification keys. However, morphological characters are sometimes difficult to be observed or do not provide a good discrimination among related taxa (Smith, 1986, Gennard, 2007, Wells & Stevens, 2010).

Flesh flies of the genera Sarcophaga (Diptera: sarcophagidae) are of considerable medical and economic importance, since they are known as myiasis producing agents in animals and humans; and they can be used to determine the post mortem interval (Gomeset. al., 2003). The development stage of insect species helps forensic specialist to determine time since death .Recovered insects from human cadavers, mostly fleshfly and flesh flies larvae, can provide information on the conditions experienced by a body following death To determine time since death, considerations of the critical factors affecting the rate of decomposition are important. These factors include location of the body, temperature, general climate, time of year, insect activity, animal activity in the area, and the amount of rainfall (Nafte, 2000). Temperature is the most important factor affecting developmental rate Temperature and access to the cadaver are two important factors affecting insect succession and temperatures generally reduce the developmental period of Diptera.(Campobasso et al., 2001).

Copyright to IJARSCT www.ijarsct.co.in DOI: 10.48175/IJARSCT-8132

# IJARSCT



#### International Journal of Advanced Research in Science, Communication and Technology (IJARSCT)

Volume 3, Issue 3, January 2023

#### **II. MATERIALS AND METHODS**

Sarcophaga Africalarvae were collected from dead dogat cholai village of Poladpur tehsilon raigad district (M.S)-India and reared in the laboratory in the rearing box by feeding daily on fresh liver of sheep and goat and water sweetened with honey.Morphological identification was done in the laboratory using the identification keys (Sukontason et al., 2003).About 80 eggs were collected in indifferentseasons (rainy, summer and winter) with the help of fine brush and 50 eggs each were reared at the laboratory conditionand the duration of different developmental stages and their morphological parameters (length, width and weight) were determined. The temperature and the humidity were recorded by Hygro-thermometer clock OPTILAB Model THC-20.

#### **III. OBSERVATION AND RESULTS**

Sarcophaga Africa, one of the fleshflies known as hairy maggot fleshfly, adult has face and cheeks with dense silvery hairs, anterior spiracle of the adult is open and proepisternal seta (stigmatic bristle) present, The larvae have tubercles hence called hairy maggot, these tubercles along the body segment are knobs encircling mostly half of lower surface, spines are round-knob turned spirally three times around the base of each tubercle; absence of hairy like structure at the base of tubercles in the caudal region; anterior spiracles always with 09 papillae and very rear 08 papillae. Life cycle duration of Sarcophaga Africain rainy season was completed in  $270 \pm 1.25$  hrs ( $11.25 \pm 0.40$  days), Table.1) when the maximum temperature was  $27.02^{\circ}$ C and the minimum temperature was  $26.4^{\circ}$ C, but in summer season when the maximum temperature was  $36.6^{\circ}$ C and the minimum temperature  $33.02^{\circ}$ C, the life cycle was completed in  $220\pm 1.17$  hrs ( $06.16.04 \pm 0.10$  days), (Table. 2)while in winter season cycle was completed in  $310 \pm 1.35$ hrs ( $12.91 \pm 0.21$  days)when themaximum and minimum temperatures were  $27.4^{\circ}$ C and  $17.2^{\circ}$ C respectively (Table. 3). Size of the different developmental stages varied from season to season; in summer season, the size of different stages was smaller than same stage in rainy season and bigger than the same stage in winter season.

Hours	Developed	Length	Width	Weight	Temperature (°C)			Humidity (%)			
	stage	(mm)	(mm)	(mg)	Max.	Min.	Avg	Max	Min	Avg	
15	Eggs	$1.2 \pm 0.08$	$0.4 \pm 0.07$	$0.29\pm0.02$	28.1	26.3	27.2	70	46	56.5	
39	1st Instar	$4.4 \pm 0.11$	$2.1 \pm 0.25$	$9.6 \pm 0.9$	28.1	26.3	27.2	70	40	55	
72	2nd Instar	$8.5 \pm 0.15$	$3 \pm 0.02$	$26.2 \pm 0.05$	28.2	26.2	27.2	75	41	58	
104	3rd Instar	$11.2 \pm 0.26$	$3.5 \pm 0.28$	$55.4 \pm 0.32$	27.6	26.4	27.00	75	41	58	
150	Prepupae	$10.6 \pm 0.14$	$4.2 \pm 0.9$	$46.4 \pm 0.05$	27.2	26.2	26.7	76	39.00	57.5	
270	Pupae	$8.2 \pm 0.36$	$3.2 \pm 0.219$	$39.5 \pm 0.13$	27.3	26.5	26.4	70	43.42	56.71	
	Adult	$8.3 \pm 0.27$	$3.6 \pm 0.11$	$32.3 \pm 0.19$	27.5	26.5	27.00	75	38	56.5	

Table 1: Duration of different life cycle stages of Sarcophaga Africain rainy season

±) Indicate SD of five values

Table 2: Duration of different life cycle stages of Sarcophaga Africain summer season

Hours	Developed	Length	Width	Weight (mg	Temperature (°C)			Humidity (%)		
	stage	(mm)	(mm)		Max	Min	Avg	Max	Min	Avg
11	Eggs	$1.00 \pm 0.05$	$0.2 \pm 0.04$	$0.25 \pm 0.01$	35.1	33.3	34.2	71	50	60.50
30	1st Instar	4.1± 0.09	$1.1 \pm 0.24$	$8.6 \pm 0.9$	35.1	33.3	34.2	71	48	59.50
62	2nd Instar	$7.5 \pm 0.13$	$2 \pm 0.02$	$22.2 \pm 0.04$	35.2	33.2	34.2	76	47	61.50
84	3rd Instar	$10.2 \pm 0.24$	$2.5 \pm 0.26$	$49.4 \pm 0.29$	36.6	34.4	35.50	76	49	62.50
130	Prepupae	9.6 ± 0.12	$3.2 \pm 0.5$	$41.4 \pm 0.2$	36.2	34.2	35.2	77	49.00	63.00
220	Pupae	$7.2 \pm 0.36$	$2.2\pm0.27$	$32.5 \pm 0.09$	36.3	34.5	35.4	72	53.42	62.71
	Adult	$7.3 \pm 0.27$	$2.4 \pm 0.10$	$27.3 \pm 0.13$	36.5	34.5	35.00	78	58	68

 $\pm$ ) Indicate SD of five values

## IJARSCT



International Journal of Advanced Research in Science, Communication and Technology (IJARSCT)

#### Volume 3, Issue 3, January 2023

Но	Developed	Length	Width	Weight	ight Temperature (°C)			Humidity (%)			
urs	rs stage (mm)	(mm)	(mg)	Max.	Min.	Avg	Max.	Min.	Avg		
20	Eggs	$1.43 \pm 0.02$	$0.4 \pm 0.05$	$0.22 \pm 0.02$	27.4	21.1	24.3	31	26	28.50	
48	1st Instar	$4.5 \pm 0.12$	$1.4 \pm 0.11$	$8.2 \pm 0.15$	23.6	20.1	22.4	31	30	30.50	
80	2nd Instar	$7.2 \pm 0.21$	$2.8\pm0.12$	$24.3\pm0.27$	23.2	21.2	22.8	29	27	28.00	
120	3rd Instar	$9.6 \pm 0.07$	$3.5 \pm 0.26$	$47.7\pm0.20$	21.6	19.2	21.1	26	25	25.50	
170	Prepupae	$8.4 \pm 0.20$	$4.4 \pm 0.21$	$47.2 \pm 0.21$	21.6	18.2	20.5	25	24	24.50	
310	Pupae	8 ± 0.31	$4. \pm 0.14$	$42.4\pm0.34$	21.8	17.6	20.1	25	23	24.00	
	Adult	8.1 ± 0.22	$3.7 \pm 0.24$	$45.1\pm0.03$	21.6	17.2	19.8	24	20	22.00	

±) Indicate SD of five values

#### **IV. DISCUSSION**

Sarcophaga Africais a species of medical and economic importance (Sukontason et al. 2008) and playing an important role in solving the forensic cases (Smith, 1986;) This flesh fly is one of the first colonizers of the corpse. Higher temperatures generally prop up egg hatching and accelerate maturation of larvae which can double their size in few hours. If the sarcophagidae larvae have reached maximum length at the peak of feeding, they tend to decline progressively and about 75% of the sarcophagidpre-adult cycle may be spent in post feeding and pupation .The morphological parameter of different stages differs from season to season. Larvae were healthy and bigger in size in rainy season but in summer season life cycle duration was short and the size of different stages was small while in winter season the life cycle duration was longer than rainy season but the size in winter season also smaller than the size in rainy season. Study on the effect of temperature on the different developmental stage of Sarcophaga Africaand life cycle duration in rainy season and low constant temperature 10 °C reported that in rainy season life cycle duration completed in 11.04 ± 0.08 days when the maximum and minimum temperature were 29°C and 26°C respectively. But in low constant temperature 10 °C life cycle was completed in  $25.38 \pm 0.16$  days (AbdAlgalil and Zambare, 2015), they reported impact of temperature on the morphological parameters in rainy season and low constant temperature. Effect of fluctuation of temperature on development of sarcophagidflies Protophormiaterraenovae was reported at 4-28°C and 9-23°C to their mean constant temperature, 16°C and, found that generally development at the greater fluctuation was fast and at the constant temperature was slow. The effect of summation rate is suspected to have caused this difference in development rate because fluctuations above the mean enhance the rate comparatively more than temperatures below the meancan lower the rate (Warren and Anderson, 2013). For forensic investigations, entomological evidences found in criminal scene around the corpse are collected and preserved according to medico-legal standard procedures. Alsomicroclimatic temperatures obtainable in the maggot's immediate environment at criminal site is established and linked retrospectively with the air temperature records. Assuming an average constant temperature, as is the case with corpses found indoors, maggots or pupae which recovered from the scene are stored at a constant temperature till they pupate or the first adults emerge out. Then their age can be used for PMI determination (Grassberge and Reiter, 2002).

#### V. CONCLUSION

In this study the effect of temperature on the life cycle of *Sarcophaga Africa* in different season indicate that life cycle duration in rainy season was completed in  $11.25 \pm 0.40$  days, but in summer season was  $6.16. \pm 0.10$  days, days while in winter season was completed in  $12.91 \pm 0.21$  days. The high temperature accelerated the development in summer and delayed the development in winter season by about 3 days. Larvae were healthy and bigger in size in rainy season and small in summer season, while in winter season larvae were smaller than summer and rainy season. Temperature plays an important role in period of life cycle stages and hence correct temperature changes should be considered for PMI determination after the life cycle stages of *Sarcophaga Africa* collected from corpse.

Copyright to IJARSCT www.ijarsct.co.in DOI: 10.48175/IJARSCT-8132

# **IJARSCT**



## International Journal of Advanced Research in Science, Communication and Technology (IJARSCT)

#### Volume 3, Issue 3, January 2023

#### REFERENCES

- [1]. Smith KGV. (1986): A Manual of Forensic Entomology. Trustees of the British Museum (natural history). London. 1986.
- [2]. Gennard D. (2007): Forensic Entomology. An Introduction. John Wiley, Sons Ltd, England. 2007.
- [3]. Wells JD, Stevens JR.(2010): Molecular methods for Forensic Entomology. In: Forensic entomology: the utility of arthropods in legal investigations 2nd edition (JH Byrd JH, Castner JL, eds.) CRC Press, Boca Raton. 2010. pp. 437-452.
- [4]. Gomes, L., Augusto, W., Godoy, C. and Zuben, C.V. (2003): A review of post feeding larval dispersal in fleshflies: implications for forensic entomology. Naturwissenschaften. 93: 207-215.
- [5]. Nafte, M. (2000): Flesh and Bone: An Introduction to Forensic Anthropology. Durham, NC: Carolina Academic Press Queiroz, M.M.C.
- [6]. Campobasso C.P., Di Vella G.andIntrona, F. (2001): Factors affecting decomposition and Diptera colonization. Forensic Sci. Int. 120: 18-27.
- [7]. Sukontason, K., Methanitikorn, R., Sukontason, K.L., Piangjai, S. and Olson, J.K.(2003): clearing technique to examine the cephalopharygeal skeletons of flesh fly larvae. J .Vector Ecol. 29: 192-195.
- [8]. Smith, K.G.V. (1986): A manual of forensic entomology, London, UK: Cornell University Press.
- [9]. Abd Algalil, F.M. and Zambare S.P. (2015): Effects of Temperature on the Development of sarcophagid fly of forensic importance Oxysarcodexia terminalis.Indian Journal of Applied Research, 5: 767-769.
- [10]. Warren, J.A. and Anderson, G.S. (2013): Effect of Fluctuating Temperatures on the Development of a Forensically Important Blow Fly, Protophormiaterraenovae (Diptera: sarcophagidae) Environmental Entomology. 42:167-172.
- [11]. Grassberger, M. and Reiter, C. (2002): Effect of temperature on development of the forensically important holarctic blow fly Protophormiaterraenovae (Robineau-Desvoidy) (Diptera: sarcophagidae) Forensic Science International.128: 177-182.