

Herbal Extracts as Antidiabetic and Antioxidant Candidates: A Narrative Review of Experimental Models, Mechanisms, Standardization and Translational Cautions

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Abstract: *Diabetes mellitus is a progressive metabolic disorder linked with chronic hyperglycemia, oxidative stress, dyslipidemia, beta-cell dysfunction and tissue injury. Herbal extracts are widely explored as potential antidiabetic and antioxidant candidates because they contain structurally diverse phytoconstituents such as phenolics, flavonoids, alkaloids, tannins, saponins and terpenoids. This review summarizes the scientific basis for evaluating herbal extracts in diabetes, with emphasis on disease burden, streptozotocin-nicotinamide modelling, oxidative stress markers, in vitro antioxidant assays, digestive enzyme inhibition, phytochemical standardization, molecular docking, ADMET prediction, toxicity assessment and histopathological confirmation. The review concludes that herbal extracts may demonstrate promising preclinical potential when their effects are consistent across glycemic, antioxidant, metabolic and tissue-level endpoints. However, clinical claims require standardized extracts, authenticated raw material, mechanistic confirmation, safety data and well-designed human studies.*

Keywords: diabetes mellitus; herbal extract; oxidative stress; streptozotocin-nicotinamide; phytochemicals; alpha-glucosidase; ADMET; histopathology

I. INTRODUCTION

Diabetes mellitus remains a major public health challenge because it affects glucose regulation, lipid metabolism, oxidative balance and vascular integrity. The increasing global prevalence of diabetes justifies continued search for safe, accessible and scientifically validated interventions [1-3].

Herbal medicines are frequently investigated in preclinical diabetes research. Their value lies not in unverified claims but in the possibility that complex phytochemical mixtures may influence multiple biological pathways. Because diabetes is a multifactorial disorder, an extract may act through glucose absorption, insulin response, antioxidant defense, lipid metabolism, inflammatory modulation or tissue protection [24-30].

A professional review of herbal antidiabetic research must therefore consider methodology as much as biological outcome. An extract without botanical authentication, dose reproducibility and phytochemical characterization cannot be reliably compared across studies. Similarly, a glucose-lowering result without oxidative and histological endpoints may not establish tissue protection.



II. LITERATURE SEARCH AND REVIEW APPROACH

This narrative review was prepared by organizing evidence around the experimental thesis framework for antidiabetic and antioxidant herbal extract evaluation. Topics included diabetes burden, STZ-nicotinamide animal modelling, oxidative stress, in vitro antioxidant assays, alpha-amylase and alpha-glucosidase inhibition, extract standardization, animal safety, histopathology and computational support by docking or ADMET prediction. Priority was given to guideline sources, experimental model papers, oxidative stress reviews, phytochemical standardization methods and preclinical pharmacology reporting guidance.

III. DIABETES PATHOPHYSIOLOGY AND OXIDATIVE STRESS

Chronic hyperglycemia damages tissue through multiple biochemical routes. Increased mitochondrial reactive oxygen species, advanced glycation end products, polyol pathway activation and protein kinase C signaling contribute to endothelial dysfunction, renal stress, neuropathy, beta-cell injury and inflammatory amplification [7-10].

Antioxidant markers are therefore central to experimental interpretation. SOD, catalase, GPx and GSH represent endogenous defense capacity, whereas MDA reflects lipid peroxidation. A meaningful antidiabetic-antioxidant extract should ideally reduce hyperglycemia and simultaneously restore antioxidant balance.

Beta cells are vulnerable to oxidative injury because their antioxidant reserves are relatively limited. This creates a strong rationale for testing extracts that combine glucose-lowering and antioxidant actions in models where beta-cell function is partially preserved.

IV. EXPERIMENTAL MODELS FOR HERBAL ANTIDIABETIC SCREENING

The STZ-nicotinamide model is commonly used to create a type-2-like diabetic state in rodents. Streptozotocin injures pancreatic beta cells, while nicotinamide partially protects them, resulting in moderate hyperglycemia with residual insulin responsiveness. This model is suitable for testing extracts that may improve glycemic control, antioxidant status and pancreatic tissue morphology [4-6].

Table 1. Experimental model considerations in herbal antidiabetic research.

Model/endpoint	Strength	Limitation
STZ-nicotinamide model	Produces partial beta-cell injury and moderate hyperglycemia	Does not fully replicate human type 2 diabetes
FBG and OGTT	Measures fasting and post-load glucose control	Can be influenced by stress, feeding and sampling
HbA1c and insulin	Reflects chronic glycemic burden and endocrine function	Requires reliable kits and sample handling
Pancreatic histology	Confirms tissue-level protection	Needs blinded scoring and representative plates

V. PHYTOCHEMICAL STANDARDIZATION

Phytochemical standardization is essential because herbal extracts vary with plant identity, plant part, collection season, geographical source, drying method, solvent ratio, extraction time and storage condition. Standardization allows biological findings to be linked to a defined test material.

Total phenolic content and total flavonoid content are widely used as batch-quality indicators. Qualitative phytochemical tests can guide interpretation, but they should not be treated as definitive proof of mechanism. Chromatographic fingerprinting by HPTLC, HPLC or LC-MS strengthens reproducibility and translational value.

Table 2. Quality attributes required for herbal extract studies.

Quality attribute	Reason for importance
Botanical authentication	Prevents invalid interpretation due to wrong plant identity
Voucher specimen	Provides traceability and future verification



Extract yield	Supports reproducible dosing
TPC/TFC values	Indicate polyphenol-rich character
Chromatographic fingerprint	Confirms marker consistency across batches
Storage conditions	Protects sensitive phytoconstituents from degradation

VI. IN VITRO ANTIOXIDANT ASSAYS

DPPH and ABTS assays estimate radical scavenging capacity through electron or hydrogen donation, while FRAP evaluates ferric reducing capacity. These assays are rapid and useful for screening, but they cannot replace biological antioxidant markers. A strong review interpretation requires linking in vitro radical scavenging with in vivo changes in SOD, catalase, GPx, GSH and MDA [21-23].

VII. DIGESTIVE ENZYME INHIBITION

Alpha-amylase and alpha-glucosidase inhibition can reduce carbohydrate digestion and delay post-prandial glucose rise. Several phytoconstituents, particularly phenolics and flavonoids, have been evaluated for this mechanism. However, enzyme inhibition in a test tube must be confirmed by OGTT and metabolic endpoints before being considered biologically meaningful [24-26].

VIII. IN VIVO BIOCHEMICAL ENDPOINTS

An integrated in vivo assessment usually includes FBG, OGTT, HbA1c, serum insulin, lipid profile, hepatic markers, renal markers, antioxidant enzymes and histopathology. A consistent direction across these endpoints strengthens the argument for extract activity. For example, reduced glucose with improved HbA1c, restored insulin, lower MDA and improved pancreatic histology is more convincing than a single fall in fasting glucose.

Table 3. Endpoint interpretation in antidiabetic-antioxidant studies.

Endpoint	Meaning
FBG	Primary indicator of fasting glycemic control
OGTT AUC	Integrated post-load glucose exposure
HbA1c	Chronic glycemic burden
Serum insulin	Endocrine support or beta-cell function
Lipid profile	Secondary metabolic benefit
SOD/CAT/GPx/GSH	Endogenous antioxidant defense
MDA	Lipid peroxidation and membrane injury
Histology score	Tissue-level confirmation

IX. ROLE OF MOLECULAR DOCKING AND ADMET PREDICTION

Computational methods can support herbal antidiabetic research by identifying plausible interactions between marker compounds and targets such as alpha-glucosidase, alpha-amylase, PPAR gamma, DPP-4, aldose reductase or antioxidant pathway-related proteins. ADMET tools help screen drug-likeness, gastrointestinal absorption, CYP risk, transporter liability and predicted toxicity [11-13]. These methods should be described as supportive and hypothesis-generating rather than confirmatory.

X. TOXICITY AND ETHICAL REPORTING

Preclinical herbal extract studies require responsible safety interpretation. Acute oral toxicity observations may support dose selection, but they do not establish chronic safety. Longer subacute and subchronic toxicity studies are necessary before translational claims are made [14,15].



Animal studies should report group allocation, number of animals, randomization, blinding where applicable, humane endpoints, statistical methods and raw data traceability. ARRIVE-style reporting improves transparency and reduces overstatement [16,17].

XI. TRANSLATIONAL LIMITATIONS

Most herbal antidiabetic studies remain preclinical. Differences between animal models and human diabetes, extract variability, unknown active constituents, incomplete toxicity data and lack of clinical pharmacokinetics limit translation. Therefore, professional claim language should state that an extract shows preclinical potential rather than clinical efficacy.

XII. FUTURE PERSPECTIVES

Future studies should combine standardized extraction, LC-MS/HPTLC fingerprinting, marker-based quantification, mechanism-specific assays, subchronic toxicity, formulation development and carefully designed clinical evaluation. Multi-omics approaches, insulin signaling assays, Nrf2/NF-kB pathway markers and beta-cell immunostaining may further clarify mechanism.

XIII. CONCLUSION

Herbal extracts can be scientifically evaluated as antidiabetic and antioxidant candidates when studies are standardized, controlled and cautiously interpreted. The strongest evidence emerges when phytochemical standardization aligns with in vitro antioxidant activity, digestive enzyme inhibition, improved glycemic endpoints, restored antioxidant enzymes, lower MDA and improved pancreatic histology. Such findings justify further investigation but not clinical claims until human evidence is available.

REFERENCES

1. American Diabetes Association Professional Practice Committee. Standards of care in diabetes. Diabetes Care. 2026.
2. Banerjee P, Eckert AO, Schrey AK, Preissner R. ProTox-II: a webserver for the prediction of toxicity of chemicals. Nucleic Acids Res. 2018;46:W257-W263.
3. Benzie IFF, Strain JJ. The ferric reducing ability of plasma as a measure of antioxidant power. Anal Biochem. 1996;239:70-76.
4. Blois MS. Antioxidant determinations by the use of a stable free radical. Nature. 1958;181:1199-1200.
5. Brownlee M. Biochemistry and molecular cell biology of diabetic complications. Nature. 2001;414:813-820.
6. Chakote VR, Dhembre GN, Rajguru JR, Joshi DA. In-Situ Gel for Nasal Drug Delivery. *International Journal of Development Research*. 2018;8(2):18763-18769
7. Daina A, Michielin O, Zoete V. SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness. Sci Rep. 2017;7:42717.
8. Dirir AM, Daou M, Yousef AF, Yousef LF. A review of alpha-glucosidase inhibitors from plants as potential candidates for diabetes therapy. Phytochem Rev. 2021;20:1049-1079.
9. Fu L, Shi S, Yi J, et al. ADMETlab 3.0: an updated comprehensive online ADMET prediction platform. Nucleic Acids Res. 2024;52:W422-W431.
10. Gheibi S, Kashfi K, Ghasemi A. A practical guide for induction of type-2 diabetes in rat: incorporating a high-fat diet and streptozotocin. Biomed Pharmacother. 2017;95:605-613.
11. Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res. 2010;107:1058-1070.
12. Harborne JB. Phytochemical Methods. London: Chapman and Hall; 1998.
13. International Diabetes Federation. IDF Diabetes Atlas. 11th ed. Brussels: International Diabetes Federation; 2025.



14. J Rajguru R. Development and Validation of RP-HPLC and UV Spectrophotometric Absorptivity Method for Simultaneous Estimation of Cyclobenzaprine Hydrochloride and Aceclofenac in Pharmaceutical Dosage Form. *International Journal of Science & Engineering Development Research*. 2020;5(4):320–329.
15. Jeevan Rajguru R. Magnetic and pH Sensitive Nanoparticles for Cancer Drug Delivery. *Asian Journal of Pharmaceutical Research and Development*. 2019;7(4).
16. JR. Rajguru Development and Evaluation of Nasal Mucoadhesive In-Situ Gel Formulations of Carbamazepine Using In-Vitro, Ex-Vivo and In-Vivo Methods. *International Research Journal of Pharmacy and Medical Sciences*. 2018;1(6).
17. Kazeem MI, Adamson JO, Ogunwande IA. Modes of inhibition of alpha-amylase and alpha-glucosidase by aqueous extract of Morinda lucida. *BMC Complement Altern Med*. 2013;13:91.
18. Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: the ARRIVE guidelines. *PLoS Biol*. 2010;8:e1000412.
19. King AJF. The use of animal models in diabetes research. *Br J Pharmacol*. 2012;166:877-894.
20. Kokate CK. Practical Pharmacognosy. New Delhi: Vallabh Prakashan; 2014.
21. Kumar S, Narwal S, Kumar V, Prakash O. Alpha-glucosidase inhibitors from plants: a natural approach to treat diabetes. *Pharmacogn Rev*. 2011;5:19-29.
22. Maritim AC, Sanders RA, Watkins JB. Diabetes, oxidative stress, and antioxidants. *J Biochem Mol Toxicol*. 2003;17:24-38.
23. Masiello P, Broca C, Gross R, et al. Experimental NIDDM: development of a new model in adult rats administered streptozotocin and nicotinamide. *Diabetes*. 1998;47:224-229.
24. OECD. Test No. 423: Acute Oral Toxicity - Acute Toxic Class Method. Paris: OECD Publishing; 2002.
25. OECD. Test No. 425: Acute Oral Toxicity - Up-and-Down Procedure. Paris: OECD Publishing; 2008.
26. Percie du Sert N, Hurst V, Ahluwalia A, et al. The ARRIVE guidelines 2.0. *PLoS Biol*. 2020;18:e3000410.
27. Radenkovic M, Stojanovic M, Prostran M. Experimental diabetes induced by streptozotocin and alloxan: methodological comparison. *J Pharmacol Toxicol Methods*. 2016;78:13-31.
28. Rajguru JR. Cancer Chemotherapy with Peptides and Peptidomimetics Drug and Peptide-Based Vaccines. *International Research Journal of Pharmacy and Medical Sciences*. 2019;2(2).
29. Re R, Pellegrini N, Proteggente A, et al. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic Biol Med*. 1999;26:1231-1237.
30. Ríos JL, Francini F, Schinella GR. Natural products for the treatment of type 2 diabetes mellitus. *Planta Med*. 2015;81:975-994.
31. Satyanarayana N, et al. Medicinal plants and phytochemicals in diabetes management: mechanisms and translational considerations. *Front Nutr*. 2022.
32. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols by Folin-Ciocalteu reagent. *Methods Enzymol*. 1999;299:152-178.
33. Szkudelski T. The mechanism of alloxan and streptozotocin action in beta cells of the rat pancreas. *Physiol Res*. 2001;50:537-546.
34. Tran N, Pham B, Le L. Bioactive compounds in anti-diabetic plants: from herbal medicine to modern drug discovery. *Biology*. 2020;9:252.
35. Unuofin JO, Lebelo SL. Antioxidant effects and mechanisms of medicinal plants and their bioactive compounds for the prevention and treatment of diabetes. *Oxid Med Cell Longev*. 2020;2020:1356893.
36. World Health Organization. Global report and fact sheet updates on diabetes. Geneva: WHO; 2024.
37. Yan LJ. Pathogenesis of chronic hyperglycemia: from reductive stress to oxidative stress. *Cells*. 2022;11:2971.

