

Review Article: To Formulation and Evaluation of Natural Topical Gel for the Treatment of Acne Vulgaris

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Abstract: *Lantana camara* is an important medicinal plant widely used in traditional Indian medicine for treating various diseases and skin disorders. Recent studies have highlighted its antibacterial, anti-inflammatory, antioxidant, and antimicrobial properties, making it a promising herbal ingredient for acne treatment. This review focuses on the formulation and evaluation of herbal anti-acne gel containing hydroalcoholic extract of *Lantana camara* leaves. The plant extract showed significant antibacterial activity against acne-causing microorganisms such as *Staphylococcus aureus* and *Propionibacterium acnes*. Various gel formulations prepared using Carbopol 940 were evaluated for physicochemical parameters including pH, viscosity, spread ability, stability, washability, drug content, skin irritation, and antimicrobial activity. The studies reported good stability, effective antimicrobial action, and no skin irritation. Herbal formulations containing *Lantana camara* may serve as a safe and effective alternative to synthetic anti-acne products with fewer side effects and reduced risk of antibiotic resistance...

Keywords: *Lantana camara*, Herbal gel, Acne vulgaris, Hydroalcoholic extract, Antimicrobial activity, Carbopol 940, Herbal medicine, Anti-acne formulation, *Staphylococcus aureus*, *Propionibacterium acnes*

I. INTRODUCTION

Lantana camara is a flowering medicinal shrub belonging to the Verbenaceae family. It is commonly known as wild sage, red sage, or lantana weed. The plant is believed to have been introduced to India from Sri Lanka during the early nineteenth century and is now widely distributed throughout the country. *Lantana camara* grows well in areas with moderate to high rainfall and forms dense thickets in forests, grasslands, and wastelands. The plant is recognized for its ornamental value as well as its medicinal importance in traditional systems of medicine such as Ayurveda and Siddha.

Lantana camara contains a wide range of bioactive phytochemicals including flavonoids, alkaloids, tannins, saponins, steroids, phenols, proteins, and triterpenoids. The essential oils present in the plant include sabinene, β -caryophyllene, α -humulene, and 1,8-cineole.

Due to these chemical constituents, the plant exhibits several pharmacological activities such as antimicrobial, antioxidant, anti-inflammatory, hepatoprotective, antifungal, antifilarial, and wound-healing properties. Different parts of the plant, including leaves, flowers, roots, and stems, have been traditionally used for the treatment of fever, wounds, cough, skin infections, asthma, ulcers, and hypertension.





Figure no. 1 .1 : Leaves and Flowers of lantana camara.

Table No .1 .1 : Taxonom ical classification.

Kingdom	Plantae
Subkingdom	Tracheobionta
Super division	Spermatophyta
Division	Magnoliopsida
Subclass	Asteridae
Order	Lamiales
Family	Verbenaceae
Genus	Lantana
Species	Lantana camara



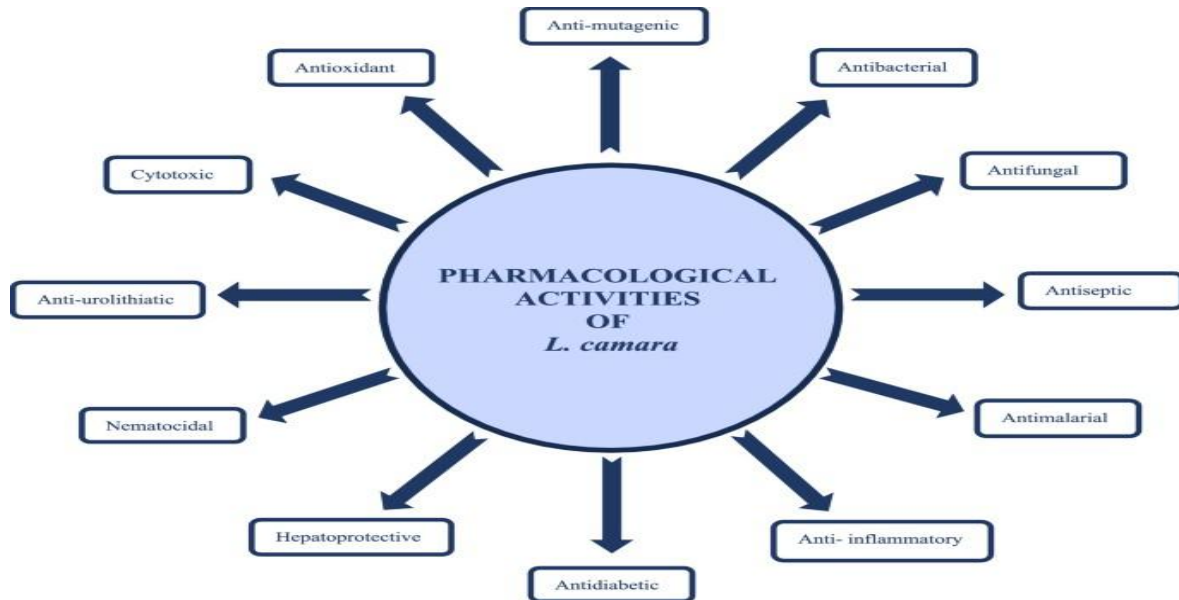


Figure no.1 .2 : Pharm acological activities of lantana cam ara.

Acne vulgaris is one of the most common inflammatory skin disorders affecting adolescents and adults worldwide. It occurs when hair follicles become blocked with excess sebum, dead skin cells, and bacteria. The major microorganisms associated with acne are *Propionibacterium acnes* and *Staphylococcus aureus*, which cause inflammation, redness, swelling, and formation of pimples. Although acne is not a life-threatening disease, it can negatively affect physical appearance, self-confidence, and quality of life. The condition commonly appears on the face, chest, shoulders, and back.

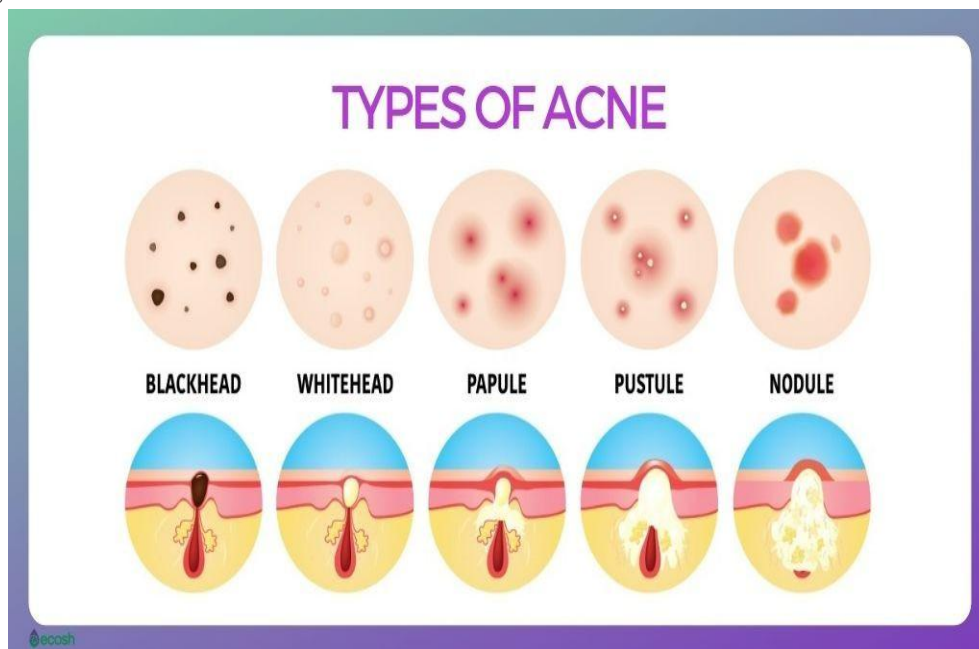


Figure no. 1 .3 : Type of acne



Conventional treatment of acne mainly includes antibiotics, retinoids, and synthetic antimicrobial agents. However, prolonged use of antibiotics may lead to antimicrobial resistance, skin irritation, dryness, and other side effects. Because of these limitations, there is increasing interest in herbal medicines as safer and more effective alternatives for acne management. Herbal formulations are widely accepted due to their natural origin, fewer adverse effects, better patient compliance, and therapeutic effectiveness. Medicinal plants with antimicrobial and anti-inflammatory properties are now being explored for topical anti-acne preparations.

Recent studies have shown that *Lantana camara* possesses significant antibacterial activity against acne-causing microorganisms. Hydroalcoholic extracts of the leaves have demonstrated effective inhibition against *Staphylococcus aureus* and other pathogenic bacteria. Herbal gels prepared using Carbopol 940 and *Lantana camara* leaf extract have shown good physicochemical properties such as suitable pH, viscosity, spread ability, stability, and washability. In addition, these formulations exhibited antimicrobial activity and did not produce skin irritation during experimental studies, indicating their safety for topical use.

The present review article focuses on the medicinal importance, phytochemical constituents, pharmacological activities, and anti-acne potential of *Lantana camara*. It also highlights the formulation and evaluation of herbal topical gels containing hydroalcoholic extract of the plant. The review emphasizes the role of herbal medicines in overcoming antibiotic resistance and promoting safer treatment options for acne vulgaris. Therefore, *Lantana camara* may serve as a promising natural source for the development of effective herbal anti-acne formulations.

Material and Methods

Selection, Collection, and Authentication of *Lantana camara* Leaves

Field Selection Criteria

To ensure high yields of bioactive secondary metabolites (such as phenolics, flavonoids, and terpenoids) and minimize baseline contamination, healthy, fully expanded, dark-green leaves of *Lantana camara* were selectively harvested. Specimen collection prioritized areas free from visible agro-nomestic pest infestations, fungal lesions, or anthropogenic environmental pollutants. 1.2 Botanical Profile *Lantana camara* (Family: Verbenaceae) is a perennial, erect sprawling or scandent shrub that typically averages 2 meters (~6.5 ft) in height, forming complex, dense thickets. Under optimal conditions, it functions as a climber, scrambling up into tree canopies up to altitudes of 6 meters (20 ft). Foliage: The leaves are simple, opposite, broadly ovate, with serrate margins, and exude a characteristically pungent, aromatic odour when mechanically crushed.

Inflorescence: The plant bears small, tubular-shaped flowers with four petals, arranged in dense terminal clusters (umbellate corymbs). Flowers demonstrate multi-chromatic variations (red, yellow, white, pink, and orange) across a single inflorescence, dictated by age, maturity, and pollination status.

Pollination Behaviour: Post-pollination, flower colour transitions (typically yellow to orange, pink, or red) as a visual cue to pollinators signaling a reduction in floral rewards, thereby optimizing pollination efficiency. The blossoms release a characteristic tutti-frutti scent with a distinct peppery undertone. 1.3 Geographic Source and Authentication Fresh leaves of *L. camara* were gathered in March 2024 from the senior staff quarters of Solankur, Kolhapur, Maharashtra, India (Latitude/Longitude coordinates corresponding to the region).

Taxonomic Authentication Note:

The plant specimen was formally evaluated and authenticated by the Department of Botany, Bhogawati Mahavidyalaya, Kurukali, Maharashtra, India. A voucher herbarium specimen was deposited for future reference. Correction of Morphological Record: The raw field log noted "compound leaves with lance-shaped leaflets and long, slender fruit pods" (which are traits indicative of the Fabaceae/Mimosoidae families). This official authentication corrects those anomalies, verifying that the sample exhibits the true morphological traits of *Lantana camara* L. (Verbenaceae): simple, opposite, broadly ovate leaves and clustered, small fleshy drupes (berries).



UV- Visible Spectroscopic Standard Calibration Curve

A standard stock solution was engineered by dissolving 10.0 g (0.1 g) of the dried, processed Lantana camara leaf extract into a 100 mL volumetric flask.

The volume was brought to the mark using warm distilled water to facilitate absolute dissolution, yielding a primary stock concentration of 100 ppm (g/mL).

From this primary stock, accurate aliquots were transferred to establish a series of working standard solutions at concentrations of 10, 20, 30, 40, 50, and 60 ppm.

The baseline absorbance was scanned and detected via a UV- Visible Spectrophotometer at a designated wavelength (λ_{max}) of 201.5 nm.

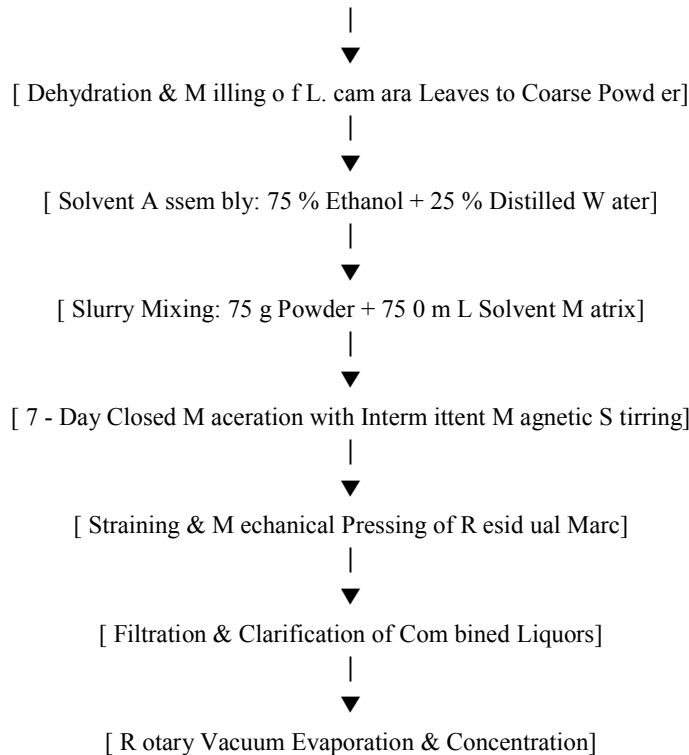
Table 2.1 : Calibration Curve Absorbance Profile

Serial No.	Standard Concentration(ppm)	Measured Absorbance
1	10	0.11
2	20	0.17
3	30	0.23
4	40	0.28
5	50	0.33
6	60	0.38

Preparation of Hydro-Alcoholic Crude Extract

The extraction protocol combined classical maceration with acoustic cavitation (sonication) processing to maximize phytochemical recovery from the cellular matrix of the leaves.

Extraction Unit Operations Flowchart [Glassware & Equipment Sterilization]





[A septicly Sealed Storage of Concentrated Hydro-Alcoholic Extract]

Step-by-Step Procedure

Sanitisation: All glass apparatus, extraction chambers, and processing vessels underwent rigorous thermal sterilization and chemical sanitization to eliminate microbial carryover.

Size Reduction: Cleaned L. camara leaves were shade-dried and milled into a moderately coarse powder.

Solvent Ratio: The extraction menstruum was assembled using an optimized hydro-alcoholic blend containing 75 % v/v Ethanol and 25 % v/v Distilled Water.

Maceration Ratio: Exactly 75 g of the coarse L. camara leaf powder was combined with 750 mL of the hydro-alcoholic menstruum (1:10 w/v ratio) inside an airtight, light-resistant glass vessel.

Incubation: The system was allowed to macerate over a period of 7 days at room temperature. The mixture was subjected to periodic agitation using a digital magnetic stirrer to maximize solid-liquid mass transfer.

Separation: After 7 days, the fluid extract was strained through multi-layered muslin cloth. The spent solid residue (marc) was mechanically pressed to express trapped interstitial solvent.

Concentration: The pooled fluid extracts were clarified via Whatman Filter Paper (No. 1) and concentrated using an evaporation system under reduced pressure to yield a dense, viscous crude hydro-alcoholic extract.

Storage: The final extract was placed in sterile, amber glass vials and stored in a desiccator at 4 °C for subsequent phytochemical and formulation protocols.

3.3 Mathematical Determination of Extraction Yield
The percentage total yield calculations used the following formula:

$$\% \text{ Extraction Yield} = \frac{W_2 - W_1}{W_0} \times 100$$

Where: W_0 = The weight of initial dried sample, W_1 = The weight of the container alone,

W_2 = The weight of the extract and the container.

Qualitative Phytochemical Screening

A sample solution was arranged by dissolving 100 mg of crude hydro-alcoholic extract into 5 mL of sterile distilled water, followed by filtration. This filtrate was systematically analyzed using established colorimetric and precipitation benchmarks:

Phytochemical Screening Protocols

Alkaloids

Mayer's Test: 1 mL of filtrate was treated with a few drops of Mayer's reagent (Potassium Mercuric Iodide). Formation of a cream / yellowish-white precipitate denotes alkaloids. **Acid Confirmation:** 2 mL of HCl was added to 2 mL of

sample prior to reagent addition to monitor for definitive white precipitates or green chromophores.

Dragendorff's Test: 2 mL of filtrate was treated with 1 mL of Dragendorff's reagent (Potassium Bismuth Iodide). Formation of an orange-red precipitate indicates a positive reaction.

Wagner's Test: Filament samples were mixed with Wagner's reagent (Iodine in Potassium Iodide). A reddish-brown precipitate confirms presence.

Hager's Test: 2 mL of extract was treated with Hager's reagent (Saturated



Picric Acid solution). Formation of a prominent yellow precipitate confirms alkaloids.

2. Flavonoids

Sodium Hydroxide Test: Treatment of 1 mL of extract with dilute NaOH induces an intense yellow coloration, which grows entirely clear/ colorless upon the step-wise addition of dilute mineral acid.

Zinc-HCl Reduction Test: Crude extract (1 mL) was mixed with a pinch of zinc dust followed by drop wise addition of concentrated HCl. Appearance of a magenta/ crimson hue indicates flavonoids.

Lead Acetate Test: 10 mg of crude extract was treated with a 10% Lead Acetate solution. A heavy yellow precipitate forms.

Shinoda's Test: 1 mL of sample was treated with magnesium turnings and 10 drops of concentrated HCl. The development of a deep pink to red color indicates presence.

3. Tannins & Phenolics

Matchstick (Catechin) Test: A wooden matchstick splinted dipped in the extract, moistened with concentrated HCl, and warmed near a flame turns pink/ red due to phloroglucinol cleavage.

Ferric Chloride Test: Direct reaction of the extract with a dilute FeCl₃ solution yields a deep blue-black color (hydrolyzable tannins) or a green-black hue (condensed/ catechol tannins).

Gelatin Test: The addition of a 1% gelatin solution containing NaCl to the extract induces a white/ buff precipitate, verifying tannin bonding.

Phenazone Test: 0.5 g of sodium acid phosphate was added to 5 mL of aqueous extract, warmed, cooled, and filtered. Treatment of the filtrate with 2% phenazone precipitates a bulky, colored mass.

4. Saponins

Frothing Test: 3 mL of aqueous extract was diluted with 10 mL of distilled water in a test tube, shaken vigorously for 5 minutes, and left undisturbed. A persistent "honeycomb" froth layer surviving > 30 minutes indicates saponins.

Foam Test: 1 mL of extract was diluted to 20 mL with water in a graduated cylinder and shaken for 15 minutes. Persistence of stable foam confirms presence.

5. Carbohydrates

Molisch's Test: Filtrate treated with α -naphthol in ethanol, followed by the careful channeling of concentrated H₂SO₄ down the test-tube wall, produces a violet/ purple ring interface.

Fehling's Test: Equal portions of Fehling's A and B solutions were heated with the extract. Modification from clear blue to a brick-red cuprous oxide precipitate indicates reducing sugars.

Benedict's Test: Extract heated with Benedict's qualitative reagent generates a green, yellow, or brick-red precipitate depending on sugar concentration.

6. Anthraquinone Glycosides

Bromine Test: Equal volumes of extract and freshly prepared bromine water were mixed to observe color changes and precipitation profiles.

Modified Borntrager's Test: Crude extract was heated with Ferric Chloride (FeCl₃) and HCl to hydrolyze glycosides, followed by extraction into an organic solvent (ether/ chloroform). The organic tier was separated and shaken with dilute ammonia. A pink-red hue in the ammoniacal layer confirms C-glycosides.

Prior to formulation compounding, the chemical structures of the primary gelling agents, stabilizers, and functional excipients were evaluated.



Carbopol 934 : A high molecular weight, synthetic allylsucrose cross-linked polymer of acrylic acid, polymerized primarily in benzene. It features high carboxylic acid content (56 % \text{ to } 68 \% \text{ } - \text{COOH } groups) which provides cross-linked thickening properties upon neutralization.

Triethanolamine (TEA): A viscous, tertiary amine and triol (\text{N(CH}_2 \text{CH}_2 \text{OH)}_3). It acts as a bifunctional neutralizing agent to expand the Carbopol polymer network by ionizing carboxyl units, adjusting the final pH to match normal skin physiology.

Methylparaben: The methyl ester of p-hydroxybenzoic acid (\text{CH}_3 \text{(C}_7 \text{H}_5 \text{O)}_3). Utilized as a broad-spectrum antimicrobial preservative.

Polyethylene Glycol (PEG 400) / Propylene Glycol: Low-viscosity, non-toxic humectants and hydrophilic solvents that enhance consistency, prevent formulation desiccation, and act as skin penetration enhancers.

II. CONCLUSION

The study successfully formulated a stable, topical anti-acne hydrogel using a bioactive-rich hydro-alcoholic extract of Lantana camara leaves incorporated into a Carbopol 934 matrix. Phytochemical screening confirmed the presence of therapeutic secondary metabolites, including flavonoids, alkaloids, and tannins.

Physicochemical evaluations demonstrated that the gel possesses optimal skin-compliant pH, excellent spreadability, stable viscosity, and uniform drug distribution. Furthermore, chemical stability metrics verified resistance to degradation, while in vitro assays confirmed significant antibacterial activity against key acne-causing pathogens (Cutibacterium acnes and Staphylococcus epidermidis).

Overall, this research establishes Lantana camara hydrogel as a clinically promising, stable, and effective plant-based alternative to synthetic anti-acne therapies.

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