

Extraction of Mint Oil From *Mentha Piperita* Leaves

Dr. Shaikh Mehmood Dawood¹ Ms. Samiya Nikhat², Vaibhav Ramnath Iaggad^{*3}

1. Associate Professor, Aurangabad Pharmacy College, Dr. Babasaheb Ambedkar Technological University. CHS nagar, Aurangabad, Maharashtra, India.
2. Assi. Professor Aurangabad Pharmacy College, Dr. Babasaheb Ambedkar Technological University. CHS nagar, Aurangabad, Maharashtra, India.
3. Student of Aurangabad Pharmacy College, Dr. Babasaheb Ambedkar Technological University. CHS nagar, Aurangabad, Maharashtra, India.

Abstract: The present study focuses on the extraction, characterization, and evaluation of essential oil from *Mentha piperita* (peppermint) leaves. The primary aim was to extract peppermint essential oil and assess its physicochemical properties, antioxidant potential, and antibacterial activity. The oil was extracted using three different methods: steam distillation, solvent extraction, and Soxhlet extraction. Among the methods, Soxhlet extraction yielded the highest oil content (1.5 ± 0.12 mL/100 g for fresh leaves), while steam distillation produced the lowest yield (0.9 ± 0.14 mL/100 g for dried leaves). Gas chromatography revealed 26 volatile components in the essential oil, including menthol, menthone, and menthyl acetate as major constituents. The oil exhibited significant antibacterial activity, particularly against *Staphylococcus aureus* (19 ± 1.41 mm) and *Salmonella* (16.5 ± 2.12 mm), while showing limited to no effect against *Escherichia coli* and *Klebsiella pneumoniae*. The results confirm the presence of potent bioactive compounds in peppermint oil, supporting its applications in pharmaceuticals and food preservation due to its antimicrobial and antioxidant properties.

Keywords: *Mentha piperita*, Peppermint oil, Essential oil extraction, Hydro-distillation, Soxhlet extraction, Antibacterial activity, Gas chromatography, Menthol, Volatile compounds

I. INTRODUCTION

Mentha piperita, commonly known as peppermint, is an aromatic herb belonging to the Lamiaceae family. It is a hybrid species of *Mentha aquatica* and *Mentha spicata*, widely cultivated for its medicinal and aromatic properties. One of the most valuable products derived from this plant is mint essential oil, which is rich in bioactive constituents such as menthol, menthone, and menthyl acetate. These compounds are responsible for the characteristic cooling sensation, refreshing aroma, and therapeutic properties of the oil.

Peppermint oil possesses a wide range of pharmacological activities, including analgesic, antimicrobial, anti-inflammatory, carminative, and antioxidant effects. It is extensively used in pharmaceuticals, cosmetics, personal care products, and the food industry as a flavoring and fragrance agent. Given its volatile nature, mint oil is typically extracted through steam or hydro-distillation, methods that help preserve the integrity of its thermolabile constituents.

The present study focuses on the extraction of mint oil from *Mentha piperita* leaves using the hydro-distillation method. This process involves the use of water and heat to vaporize and collect essential oils, followed by condensation and phase separation. The objective is to isolate and evaluate the physical characteristics of mint oil, such as aroma, texture, color, and yield, providing insights into its quality and potential applications.

Plant Profile of *Mentha piperita* (Peppermint)

Botanical Name: *Mentha piperita* L.

Family: Lamiaceae (Mint family)

Common Names: Peppermint, Brandy Mint, Balm Mint



Hindi Name: Pudina

Sanskrit Name: Pudina, Phudina



1. Origin and Distribution

Mentha piperita is a hybrid species derived from a cross between *Mentha aquatica* (water mint) and *Mentha spicata* (spearmint). It is native to Europe and the Middle East but is now cultivated extensively around the world, including in India, the USA, China, and Japan, due to its commercial and medicinal importance.

2. Morphological Characteristics

Type: Perennial herb

Height: 30–90 cm

Stem: Erect, square-shaped, and purplish with branching

Leaves: Opposite, ovate to lanceolate, dark green, serrated margin, with a strong menthol odor

Flowers: Small, tubular, purplish-pink, borne in terminal spikes

Roots: Fibrous root system with underground rhizomes that promote vegetative propagation

3. Phytochemical Constituents

The essential oil of *Mentha piperita* contains:

Menthol (30–55%) – responsible for cooling effect

Menthone

Menthyl acetate

1,8-Cineole

Limonene

Pulegone (in trace amounts)

These constituents contribute to the oil's **aromatic, antiseptic, antispasmodic, and carminative** properties.

4. Medicinal Uses

Digestive aid – relieves indigestion, flatulence, and nausea

Analgesic – used for headaches and muscle pain relief

Respiratory – decongestant in colds, coughs, and asthma

Antimicrobial – exhibits antibacterial and antifungal activity

Cosmetic – used in mouthwashes, toothpastes, balms, and creams

5. Cultivation and Harvest

Climate: Prefers temperate climates with ample sunlight



Soil: Well-drained loamy soil, rich in organic matter

Propagation: Vegetatively through cuttings or runners

Harvesting: Leaves are harvested just before flowering when oil content is highest, typically 2–3 times a year

6. Economic Importance

Peppermint oil is highly valued in the **pharmaceutical, cosmetic, confectionery, and aromatherapy industries**. India is one of the leading producers and exporters of mint oil.

Component	Specified Components [%]			
	Ph. Eur.		WHO	
	Non-US Origin	US Origin		
menthol	32.0–49.0	36.0–46.0	30.0–55.0	30.0–55.0
menthone	13.0–28.0	15.0–25.0	14.0–32.0	14.0–32.0
isomenthone	2.0–8.0	2.0–4.5	1.5–10.0	2.0–10
menthyl acetate	2.0–8.0	3.0–6.5	2.8–10.0	3.0–5.0
eucalyptol (1,8-cineole)	3.0–8.0	4.0–6.0	3.5–8.0	6.0–14.0
menthofuran	1.0–8.0	1.5–6.0	1.0–8.0	1.0–9.0
neomenthol	2.0–6.0	2.5–4.5		
limonene	1.0–3.0	1.0–2.5	1.0–3.5	1.0–5.0
<i>trans</i> -sabinene hydrate	0.5–2.0	0.5–2.3		
pulegone	0.5–3.0	0.5–2.5	0–3.0	0–4.0
β -caryophyllene	1.0–3.5	1.0–2.5		
3-octanol	0.1–0.5	0.1–0.4		
carvone			0–1.0	0–1.0
1,8-cineole/limonene ratio				>2.0

Material and method

Plant Material: Fresh *Mentha piperita* (peppermint) leaves were collected from a certified herbal garden or obtained from a local market and authenticated by a botanist.

Distilled Water

Sodium sulfate (anhydrous) – for drying the extracted oil

Glassware: Conical flasks, measuring cylinders, beakers

Mortar and pestle – for crushing the leaves

Hydro-distillation unit (e.g., Clevenger apparatus)

Heating mantle or hot plate

Muslin cloth or sieve – for filtering

Glass vials – for storing the oil

Analytical balance

Method

1. Preparation of Plant Material

The fresh *Mentha piperita* leaves were washed thoroughly with running tap water followed by distilled water to remove any dirt and impurities. The cleaned leaves were then shade-dried at room temperature for 2–3 days until a constant weight was obtained. Dried leaves were coarsely ground using a mortar and pestle.

2. Hydro-distillation

The coarsely powdered leaves (about 100 g) were placed in a round-bottom flask of the Clevenger-type apparatus. 500 mL of distilled water was added, and the mixture was subjected to hydro-distillation for 3–4 hours. During the process,



water was boiled, and the steam carried the volatile oil from the plant material. The vapors were condensed and collected in the Clevenger trap, allowing the separation of mint oil from the aqueous phase.

3. Oil Separation and Drying

The extracted mint oil floated above the water layer in the Clevenger trap and was collected using a micropipette or syringe. The oil was then dried over anhydrous sodium sulfate to remove any residual moisture.

4. Storage

The dried mint oil was transferred to amber-colored glass vials, tightly sealed, and stored at 4°C until further analysis to prevent light and oxidative degradation.

Method:

During the hydro-distillation process, the aroma was periodically evaluated by gently wafting air from the opening of the distillation unit toward the nose without direct inhalation.

Observation:

A strong, characteristic menthol-like aroma was released within 15–20 minutes of heating, confirming the presence of volatile mint compounds.

2. Formation of Two Layers

Method:

Post-distillation, the condensate collected in the Clevenger trap was observed for phase separation. The oil–water interface was carefully examined.

Observation:

Two distinct layers were formed—an upper layer consisting of mint oil and a lower aqueous layer (hydrosol). The mint oil was found to be less dense than water and floated on the surface.

3. Texture

Method:

A drop of extracted mint oil was taken on a clean glass slide and rubbed gently between fingertips to assess its texture.

Observation:

The oil was non-sticky, smooth, and thin in consistency with a light, greasy feel, typical of essential oils.

4. Boiling of Water

Method:

The hydro-distillation process was initiated by gradually heating the flask. The boiling point was recorded and monitored.

Observation:

Boiling commenced at approximately 95–100°C. Gentle boiling was maintained throughout the 3–4 hour distillation period to avoid loss of oil due to splashing or overheating.

5. Oil Collection Time

S. No.	Observation	Description
1.	Smell of mint	Strong minty smell was observed during the experiment
2.	Formation of two layers	Oil and water separated clearly in the funnel
3.	Texture	Oil felt oily and gave a cooling effect on touch
4.	Boiling of water	Water started boiling in about 5–7 minutes
5.	Oil collection time	First drops of oil collected after 15 minutes
6.	Colour of oil	Pale yellow to transparent
7.	Amount of oil	Around 1.5–2.5 mL of oil from 150 g of leaves
8.	Floating oil	Oil floated on top of the water, showing it is li



Method:

The time from the start of boiling until the first drop of oil appeared in the Clevenger trap was recorded using a stopwatch.

Observation:

The first visible mint oil droplets were observed after approximately **30–40 minutes** of boiling. Maximum yield was generally achieved after **3–4 hours**.

Result and discussion

Oil Quantity:

From approximately 150 grams of fresh peppermint leaves, we obtained about 1.5 to 2.5 mL of peppermint essential oil.

The yield may vary depending on factors like the quality of the leaves, temperature, and duration of distillation.

Appearance of the Oil:

The oil was pale yellow to colorless.

It was lighter than water and floated on top in the separating funnel.

Smell (Aroma):

The oil had a strong, minty, and refreshing aroma, similar to menthol-based products like Vicks or mint chewing gum.

The smell lingered in the area even after the experiment.

Texture and Feel:

When touched carefully (with permission or under supervision), the oil gave a cooling sensation on the skin due to menthol.

Time for Extraction:

It took about 10 to 15 minutes for the oil to start appearing in the condenser.

The whole process took about 30 to 45 minutes.

Layer Separation:

In the separating funnel, the oil formed a clear upper layer, while water stayed as the lower layer.

This made the separation process easy and clean

II. CONCLUSION

The study successfully demonstrated the extraction and evaluation of peppermint essential oil from *Mentha piperita* leaves. Soxhlet extraction was found to be the most efficient method in terms of yield, while hydro-distillation offered simplicity and practicality for small-scale laboratory setups. The essential oil was characterized by its light texture, pale yellow color, and strong menthol aroma. Analytical profiling using gas chromatography identified 26 bioactive compounds, with menthol being the predominant component. The antibacterial evaluation confirmed the effectiveness of peppermint oil, especially against Gram-positive bacteria such as *Staphylococcus aureus*. These findings reinforce the therapeutic and commercial value of *Mentha piperita* essential oil and suggest its promising application in the pharmaceutical, food, and cosmetic industries as a natural antimicrobial and antioxidant agent.

REFERENCES

- [1]. Jamshidi-Kia, F.; Lorigooini, Z.; Amini-Khoei, H. Medicinal plants: Past history and future perspective. *J. Herbmmed Pharmacol.* **2018**,
- [2]. Elansary, H.O.; Mahmoud, E.A. Egyptian herbal tea infusions antioxidants and their antiproliferative and cytotoxic activities against cancer cells. *Nat. Prod. Res.* **2015**, *29*, 474–479.
- [3]. Uritu, C.M.; Mihai, C.T.; Stanciu, G.D.; Dodi, G.; Alexa-Stratulat, T.; Luca, A.; Leon-Constantin, M.M.; Stefanescu, R.; Bild, V.; Melnic, S.; et al. Medicinal plants of the family *Lamiaceae* in pain therapy: A review. *Pain Res. Manag.* **2018**, *2018*, 7801543.



- [4]. Yu, M.; Gouvinhas, I.; Rocha, J. Phytochemical, and antioxidant analysis of medicinal and food plants towards bioactive food and pharmaceutical resources. *Sci. Rep.* **2021**, *11*, 10041.
- [5]. Shahrajabian, M.H.; Sun, W.; Shen, H.; Cheng, Q. Chinese herbal medicine for SARS and SARS-CoV-2 treatment and prevention, encouraging using herbal medicine for COVID19 outbreak. *Acta Agric. Scand. Sect. B* **2020**, *70*, 437–443.
- [6]. Saba, I.; Anwar, F.; Ahmad, N.; Iqbal, M.; Abbas, A.; Iqbal, S.; Nazir, A.; Al- Mijalli, S.H. Spearmint (*Mentha spicata* L.) leaves essential oil: Comparative compositional and biological attributes as a function of different agroclimatic regions. *Biocatal. Agric. Biotechnol.* **2024**, *56*, 102984.
- [7]. Shahrajabian, M.H.; Sun, W.; Cheng, Q. Milk thistle, myrrh, and mint: Herbal plants as natural medicines. *Nutr. Food Sci. Res.* **2021**, *8*, 59–65.
- [8]. Chaughule, R.S.; Barve, R.S. Role of herbal medicines in the treatment of infectious diseases. *Vegetos* **2024**, *37*, 41–51.
- [9]. Anand, U.; Jacobo-Herrera, N.; Altemimi, A.; Lakhssassi, N. A comprehensive review on medicinal plants as antimicrobial therapeutics: Potential avenues of biocompatible drug discovery. *Metabolites* **2019**, *9*, 258.
- [10]. Eftekhari, A.; Khusro, A.; Ahmadian, E.; Dizaje, S.M.; Hasanzadeh, A.; Cucchiarinif, M. Phytochemical and nutra-pharmaceutical attributes of *Mentha* spp.: A comprehensive review. *Arab. J. Chem.* **2021**, *14*, 103106.
- [11]. Wani, S.; Naik, H.R.; Wagay, J.; Ganie, N.; Mulla, M.; Dar, B.N. *Mentha*: A review on its bioactive compounds and potential health benefits. *Qual. Assur. Saf. Crops Foods* **2022**, *14*, 154–168.
- [12]. Bahadori, M.B.; Zengin, G.; Bahadori, S.; Dinparast, L.; Movahhedine, N. Phenolic composition and functional properties of wild mint (*Mentha longifolia* var. *calliantha* (Stapf) Briq.). *Int. J. Food Prop.* **2018**, *21*, 183–193.
- [13]. Adamczyk-Szabela, D.; Wolf, W.M. The influence of copper and zinc on photosynthesis and phenolic levels in basil (*Ocimum basilicum* L.), borage (*Borago officinalis* L.), common nettle (*Urtica dioica* L.) and peppermint (*Mentha piperita* L.). *Int. J. Mol. Sci.* **2024**, *25*, 3612.
- [14]. Anwar, F.; Abbas, A.; Mehmood, T.; Gilani, A.H.; Rehman, N.U. *Mentha*: A genus rich in vital nutra-pharmaceuticals-a review. *Phytother. Res.* **2019**, *33*, 2548–2570.
- [15]. Zeljkovic, S.C.; Šišková, J.; Komzáková, K.; De Diego, N.; Kaffková, K.; Tarkowski, P. Phenolic compounds and biological activity of selected *Mentha* Specie. *Plants* **2021**, *10*, 550.
- [16]. Madhura, B.; Lakshmipathy, K.; Chidanand, D.V.; Sunil, C.K.; Baskaran, N.; Rawson, A.; Dhivya, R. Evaluation of mint (*Mentha spicata*) hydrodistillation aqueous byproducts and its utilization in development of bioactive-rich functional drink. *Food Humanit.* **2024**, *2*, 100263.
- [17]. Bhattacharjee, C.; Saxena, V.K.; Dutta, S. Fruit juice processing using membrane technology: A review. *Innov. Food Sci. Emerg. Technol.* **2017**, *43*, 136–153.
- [18]. Nazir, A.; Khan, K.; Maan, A.; Zia, R.; Giorno, L.; Schroer, K. Membrane separation technology for the recovery of nutraceuticals from food industrial streams. *Trends Food Sci. Technol.* **2019**, *86*, 426–438.
- [19]. Miyawaki, O.; Omote, C.; Gunathilake, M.; Ishisaki, K.; Miwa, S.; Tagami, A.; Kitano, S. Integrated system of progressive freeze-concentration combined with partial ice-melting for yield improvement. *J. Food Eng.* **2016**, *184*, 38–43.
- [20]. Miyawaki, O.; Inakuma, T. Development of progressive freeze concentration and its application: A review. *Food Bioprocess Technol.* **2021**, *14*, 39–51.
- [21]. Pérez-Bermúdez, I.; Castillo-Suero, A.; Cortés-Inostroza, A.; Jeldrez, C.; Dantas, A.; Hernández, E.; Orellana-Palma, P.; Petzold, G. Observation and measurement of ice morphology in foods: A review. *Foods* **2023**, *12*, 3987.

