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The Gut-Liver Axis in NAFLD Progression: Insights into Pathogenesis and Therapeutic Opportunities

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Abstract: Non-alcoholic fatty liver disease (NAFLD) represents a complex disorder characterized by hepatic lipid accumulation and inflammation, ranging from simple steatosis to non-alcoholic steatohepatitis (NASH) and fibrosis. Emerging evidence suggests that dysregulation of the gut-liver axis plays a pivotal role in the pathogenesis and progression of NAFLD. This review comprehensively examines the bidirectional communication between the gut and liver, encompassing intestinal barrier dysfunction, gut microbiota dysbiosis, bile acid metabolism, and immune-mediated responses. We explore the mechanisms by which gut-derived factors, including microbial metabolites, lipopolysaccharides (LPS), and bile acids, influence hepatic lipid metabolism, inflammation, and fibrosis in NAFLD. Furthermore, we discuss therapeutic strategies targeting the gut-liver axis, including prebiotics, probiotics, bile acid modulators, and gut barrier enhancers, with the potential to attenuate NAFLD progression. Understanding the intricate interplay between the gut and liver in NAFLD pathogenesis offers novel insights into disease mechanisms and therapeutic opportunities for the effective management of this increasingly prevalent liver disorder. Nonalcoholic fatty liver disease (NAFLD) is a complex disorder that is characterized by the accumulation of fat and inflammation in the liver. This can range from simple steatosis to more severe conditions such as nonalcoholic steatohepatitis (NASH) and fibrosis. Research has shown that the gut-liver axis, which includes the communication between the gut and liver, plays a significant role in the development and progression of NAFLD. This comprehensive review examines the two-way communication between the gut and liver, including intestinal barrier dysfunction, gut microbiota dysbiosis, bile acid metabolism, and immunemediated responses. We explore how gut-derived factors such as microbial metabolites, lipopolysaccharides (LPS), and bile acids can affect hepatic lipidmetabolism, inflammation, and fibrosis in NAFLD. Additionally, we discuss various therapeutic strategies, such as prebiotics, probiotics, bile acid modulators, and gut barrier enhancers, that target the gut-liver axis and potentially reduce the progression of NAFLD. Understanding the complex relationship between the gut and liver in NAFLD pathogenesis can provide new insights into the disease's mechanisms and offer therapeutic opportunities for managing this increasingly prevalent liver disorder.

Keywords: Non-alcoholic fatty liver disease

I. INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) has escalated into a pressing global health concern, affecting a substantial portion of the population worldwide (1). Despite its pervasive prevalence, effective therapeutic interventions for NAFLD remain scarce, prompting an urgent quest for a comprehensive understanding of its pathogenesis and the discovery of novel treatment modalities. Amidst this pursuit, the gut-liver axis has emerged as a focal point, representing a dynamic interplay between the intestinal microbiota, gut epithelium, and liver, with profound implications for NAFLD progression (2). In this comprehensive review, we explore the intricate bidirectional communication between the gut and liver in the context of NAFLD pathogenesis (3). Our examination delves into the multifaceted mechanisms underlying gut dysbiosis, intestinal barrier dysfunction, bile acid metabolism, and immunemediated responses, all of which converge to orchestrate the progression of NAFLD (4).

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Furthermore, we navigate the landscape of emerging therapeutic strategies targeting the gut- liver axis, which is poised to revolutionize the management of NAFLD potentially (5). From probiotics and prebiotics to bile acid modulators and gut barrier enhancers, these innovative interventions hold promise in reshaping the treatment paradigm for NAFLD and mitigating its associated morbidity and mortality (6). This review endeavours to provide a comprehensive synthesis of current knowledge and foster future advancements in the field by illuminating the complexities of the gut-liver axis in NAFLD pathogenesis and therapeutics (7). Through a deeper understanding of these intricate interactions, we aspire to pave the way toward more effective and personalized strategies for managing NAFLD, ultimately alleviating the burden of this burgeoning global health crisis (8).

The connection between the gut and the liver in the development of non-alcoholic fatty liverdisease (NAFLD):

Intestinal Barrier Dysfunction: Non-alcoholic fatty liver disease (NAFLD) is a complex disorder with a multifactorial etiology, and emerging evidence suggests that disruption of the intestinal barrier plays a crucial role in its pathogenesis (9). The intestinal barrier, consisting primarily of epithelial cells held together by tight junction proteins, is an essential defense mechanism against the translocation of harmful substances from the gut lumen into the systemic circulation. Non-alcoholic fatty liver disease (NAFLD) is a complex disorder with multiple causes (10). Recent evidence indicates that the breakdown of the intestinal barrier is a critical factor in its development. The intestinal barrier, mainly made up of epithelial cells held together by tight junction proteins, is an essential defense against the movement of harmful substances from the intestinal space into the bloodstream (11). In this discussion, we explore how changes in tight junction proteins, increased intestinal permeability, and the movement ofmicrobial products contribute to the breakdown of intestinal barrier integrity in NAFLD (12). Here, we discuss how alterations in tight junction proteins, increased gut permeability, and microbial product translocation contribute to intestinal barrier integrity disruption in NAFLD.

Alterations in Tight Junction Proteins: Tight junctions are multiprotein complexes that regulate the paracellular permeability of the intestinal epithelium. Essential tight junction proteins include occludin, claudins, and zonula occludens (ZO) (13). InNAFLD, dysregulation of these proteins has irregularities in these proteins have been observed, leading to a compromised barrier function (14). Studies have reported expression and abnormal distribution of tight junction proteins in both animal models and human patients with NAFLD. For example, decreased expression of occludin and ZO-1 has been seen in the intestines of NAFLD patients, indicating impaired barrier integrity.

1.1 Gut Microbiota Dysbiosis in NAFLD

Non-alcoholic fatty liver disease (NAFLD) is closely linked to alterations in gut microbiotacomposition and diversity, a phenomenon commonly referred to as dysbiosis. Dysbiosis inNAFLD encompasses changes in the relative abundance of specific microbial taxa, disruptions in microbial community structure, and alterations in functional pathways (14). In this review, we will discuss the alterations observed in gut microbial composition and diversity in NAFLD, emphasizing the role of dysbiosis in promoting hepatic inflammation, insulin resistance, and fibrosis (15).

Increased Intestinal Permeability: Disruption of tight junctions leads to increased intestinal permeability, allowing the passage of substances from the gut lumen, such asbacterial products and microbial metabolites, across the epithelial barrier (16). Variousmethods, including in vivo permeability assays and measurement of serum markers likezonulin, have shown increased gut permeability in individuals with NAFLD compared to healthy controls. This heightened permeability enables harmful substances to enter the systemic circulation, triggering inflammatory responses in distant organs, includingthe liver (17).

Movement of Microbial Products: One consequence of increased gut permeability in NAFLD is the movement of microbial products, such as lipopolysaccharides (LPS) from Gram-negative bacteria, into the portal circulation (18). Elevated levels of circulating LPS have been observed in NAFLD patients and are linked to liver inflammation and disease progression. Additionally, microbial DNA and metabolites can also move from the gut to the liver, worsening hepatic inflammation and promotingthe development of steatosis and fibrosis (19). They have been observed, leading to compromised barrier function. Studies have reported decreased expression and abnormal distribution of tight junction proteins in both animal models and human patients with NAFLD. For instance, reduced expression of occludin and ZO-1 has been documented in the intestines of NAFLD patients, indicating impaired barrier integration.

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II. PATHOPHYSIOLOGY OF NAFLD AND BILE ACID HOMEOSTASIS

In addition to being essential for the digestion and absorption of dietary fat, bile acids also function as signalling molecules that control the metabolism of fat, glucose, and energy. An important role for dysregulation of bile acid homeostasis in the pathophysiology of non-alcoholic fatty liver disease (NAFLD) is becoming more widely acknowledged (21). This section examines the roles that changes in bile acid production, secretion, and signalling pathways play in hepatic lipid metabolism and inflammation in non-alcoholic fatty liver disease (NAFLD) (22).

2.1 Modifications in Bile Acid Combination:

Bile acids are combined in the liver from cholesterol through two essential pathways: the traditional (or impartial) pathway and the other option (or acidic) pathway (23). The old style pathway, started by the compound cholesterol 7α-hydroxylase (CYP7A1), is the dominating course for bile corrosive combination. In NAFLD, hepatic articulation of CYP7A1 is frequently downregulated, prompting diminished bile corrosive union (24). This change can bring about an amassing of cholesterol and different lipids in the liver, adding to steatosis.

2.2 Disturbance in Bile Acid Discharge:

Bile acids are discharged into the bile canaliculi and put away in the gallbladder, from where they are delivered into the digestive tract upon food consumption (25). In NAFLD, hepatic steatosis and irritation can impede bile corrosive discharge. The gathering of bile acids in hepatocytes because of debilitated emission can actuate cell stress and apoptosis, fueling liver injury (26). Moreover, decreased bile corrosive emission into the digestive system can upset the enterohepatic dissemination, prompting further metabolic unsettling influences.

2.3 Bile Acid Flagging/Signaling Pathways:

Bile acids capability as flagging atoms through their communications with atomic receptors, for example, the farnesoid X receptor (FXR), and cell surface receptors, similar to the G protein-coupled bile corrosive receptor (TGR5). These receptors assume vital parts in directing bile corrosive blend, lipid digestion, and irritation (27).

FXR Flagging:

FXR is profoundly communicated in the liver and digestive tract and directs the statementof qualities engaged with bile corrosive combination, transport, and digestion (28). Initiation of FXR decreases hepatic lipogenesis and increments unsaturated fat oxidation, hence safeguarding against liver steatosis. In NAFLD, FXR flagging is in many cases disabled, prompting expanded lipogenesis, diminished unsaturated fat oxidation, and hepatic lipid aggregation (29). Also, FXR initiation has calming impacts by repressing therecord of supportive of incendiary cytokines (30).

TGR5 Flagging:

TGR5, communicated in different tissues including the liver and digestion tracts, adjustsenergy consumption and fiery reactions. Enactment of TGR5 advances the arrival of glucagon-like peptide-1 (GLP-1) from enteroendocrine cells, which upgrades insulin awareness and glucose digestion (31). In NAFLD, TGR5 flagging is disturbed, adding toinsulin opposition and ongoing irritation.

Influence on Hepatic Lipid Digestion/Metabolism:

Dysregulated bile corrosive digestion impacts hepatic lipid digestion through numerous components (32). Diminished FXR action prompts diminished articulation of littleheterodimer accomplice (SHP), a repressor of sterol administrative component restricting protein-1c (SREBP-1c), a key record factor advancing lipogenesis (33). Thus, expanded SREBP-1c movement improves all over again lipogenesis, adding to hepatic steatosis. Moreover, debilitated bile corrosive flagging disturbs mitochondrial capability and unsaturated fat oxidation, further fueling lipid aggregation in the liver (34).





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Job in Hepatic Aggravation/Inflammation:

Bile acids can enact fiery pathways in the liver through immediate and aberrant systems. Amassed bile acids can instigate endoplasmic reticulum (trama center) stress and oxidative pressure, prompting the actuation of supportive of incendiary flagging fountains, for example, the atomic element kappa-light-chain-enhancer of initiated B cells (NF-κB) pathway (35). Also, dysregulated bile corrosive motioning through FXR and TGR5 can bring about the development of favorable to fiery cytokines, advancing hepatic irritation and fibrosis (36).

III. IMMUNE-MEDIATED RESPONSES IN NAFLD: CROSSTALK BETWEEN THE GUT AND LIVER

Non-alcoholic fatty liver illness (NAFLD) movement is complicatedly connected to resistant interceded reactions that include complex associations between the stomach and liver (37). Safe cells and cytokines assume a urgent part in this crosstalk, adding to hepatic irritation, insulin obstruction, and fibrosis (38). This segment investigates the job of resistant cells and cytokines in NAFLD pathogenesis, with an emphasis on the fiery flagging pathways that driveillness movement (39).

3.1 Immune Cells in NAFLD

Kupffer Cells:

Kupffer cells, the liver-occupant macrophages, are fundamental to the safe reaction in NAFLD. They perceive and phagocytose stomach determined microbial items, for example, lipopolysaccharides (LPS), that move into the entryway course because of expanded stomach penetrability (40). Enacted Kupffer cells produce supportive of fierycytokines (e.g., TNF-α, IL-6) and receptive oxygen species (ROS), starting and sustaining hepatic irritation (41).

Invading Monocytes/Macrophages:

In NAFLD, coursing monocytes are selected to the liver, where they separate into macrophages (42). These invading macrophages, alongside Kupffer cells, add to the incendiary milieu by delivering cytokines and chemokines, worsening liver injury andadvancing fibrosis (43).

Neutrophils:

Neutrophils are early responders to hepatic irritation. They are enlisted to the liver because of chemokines, like IL-8. Neutrophils discharge myeloperoxidase (MPO) and neutrophil extracellular snares (NETs), which can incite hepatocyte harm and further irritation (44).

White blood cells/T Cells:

Both CD4+ and CD8+ White blood cells are engaged with NAFLD movement. CD4+T assistant (Th) cells, especially Th1 and Th17 subsets, produce supportive of fiery cytokines (e.g., IFN-γ, IL-17) that compound liver aggravation (45). CD8+ cytotoxic Lymphocytes can straightforwardly incite hepatocyte apoptosis and add to liver harm (46).

Natural Killer (NK) Cells and Natural Killer T (NKT) Cells:

NK and NKT cells assume double parts in NAFLD. While they can have mitigating impacts under specific circumstances, their enactment frequently brings about the development of favorable to provocative cytokines and cytotoxic particles, adding to liver injury and fibrosis (47).

3.2 Cytokines in NAFLD

Growth Putrefaction Component α (TNF-α):

TNF- α is a key supportive of fiery cytokine delivered by Kupffer cells, macrophages, and other safe cells. It enacts NF- κ B flagging, prompting the record of qualities associated with irritation and apoptosis (48). TNF- α additionally advances insulin opposition by obstructing insulin flagging pathways.

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Interleukin-6 (IL-6):

IL-6 is one more basic cytokine in NAFLD, created by Kupffer cells, hepatocytes, and other safe cells (49). It initiates the JAK/STAT3 pathway, adding to hepatic irritation, insulin opposition, and hepatocyte endurance. Raised IL-6 levels are related with the seriousness of liver irritation and fibrosis (50).

Interleukin-1β (IL-1β):

IL-1 β is delivered by enacted macrophages and assumes a critical part in liver irritation. It advances the enactment of hepatic stellate cells (HSCs), which are central participants in liver fibrosis (51). IL-1 β additionally upgrades the enrollment and initiation of otherresistant cells, enhancing the incendiary reaction.

Interleukin-17 (IL-17):

Delivered by Th17 cells, IL-17 adds to NAFLD movement by advancing the enrollment of neutrophils and macrophages to the liver (53). IL-17 likewise initiates the creation of other favorable to incendiary cytokines and chemokines, sustaining the fiery outpouring (52).

Chemokines:

Chemokines like MCP-1 (CCL2) and IL-8 (CXCL8) assume fundamental parts in enlisting monocytes, macrophages, and neutrophils to the liver. These chemokines are upregulated in NAFLD and correspond with the seriousness of liver aggravation and fibrosis (53).

3.3 Fiery/Inflammatory Signaling Pathways

NF-κB Pathway:

The NF-κB pathway is a focal controller of irritation in NAFLD. Actuation of NF-κB in light of gut-derived LPS and different improvements prompts the record of supportive of fiery cytokines, chemokines, and attachment particles (54). This advances invulnerable cell enlistment and initiation, sustaining hepatic irritation and injury.

JAK/STAT Pathway:

The JAK/STAT pathway is enacted by cytokines, for example, IL-6 and assumes a urgent part in intervening fiery reactions and hepatocyte endurance. Dysregulation of this pathway in NAFLD adds to insulin opposition, steatosis, and fibrosis (55).

Inflammasome Actuation:

Inflammasomes, especially the NLRP3 inflammasome, are actuated in light of microbialitems and metabolic pressure. Initiation of the NLRP3 inflammasome prompts the cleavage and actuation of favorable to provocative cytokines IL-1 β and IL-18, driving liver irritation and fibrosis (56).

IV. THERAPEUTIC STRATEGIES TARGETING THE GUT-LIVER AXIS: PROBIOTICS AND PREBIOTICS INNAFLD

The gut-liver axis plays a critical role in the development of non-alcoholic fatty liver disease (NAFLD), and modifying this axis represents a promising therapeutic approach. Probiotics and prebiotics have gained significant attention for their potential to alter gut microbiota, enhanceintestinal barrier function, and reduce hepatic inflammation (57). This section evaluates the effectiveness of probiotics and prebiotics in managing NAFLD.

4.1 Probiotics in NAFLD

Modifying Gut Microbial Composition: Probiotics are live microorganisms that providehealth benefits to the
host when administered in adequate amounts. Common probiotic strains include Lactobacillus,
Bifidobacterium, and Saccharomyces species (58). These probiotics can change the gut microbial composition
by increasing the presence of beneficial bacteria and decreasing pathogenic bacteria. Studies have

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demonstrated that probiotics can restore microbial balance in NAFLD patients, leading to improved gut health and metabolic function (59).

- Enhancing Intestinal Barrier Function: Probiotics improve intestinal barrier integrity by increasing the expression of tight junction proteins (e.g., occludin, claudins, ZO-1) and reducing gut permeability (60). This prevents the passage of harmful microbial products, such as lipopolysaccharides (LPS), into the portal circulation. Enhanced barrier function reduces systemic and hepatic inflammation, crucial in managing NAFLD (61).
- Alleviating Hepatic Inflammation: Probiotics exert anti-inflammatory effects by modulating immune responses and cytokine production. They can decrease pro-inflammatory cytokines (e.g., TNF-α, IL-6) and increase anti-inflammatory cytokines (e.g., IL-10) (62). Probiotics also inhibit the activation of inflammatory signaling pathways, such as the NF-κB pathway, thereby reducing hepatic inflammation and preventing liver damage.
- Clinical Evidence: Several clinical trials have investigated the effects of probiotics in NAFLD patients. For
 example, a study involving NAFLD patients receiving Lactobacillus and Bifidobacterium supplements reported
 significant reductions in liver enzymes (ALT, AST), hepatic steatosis, and inflammatory markers. Another
 study demonstrated that probiotic supplementation improved NAFLD patients' insulin sensitivity and lipid
 profiles (63)

4.2 Prebiotics in NAFLD

- **Promoting the Growth of Beneficial Bacteria**: Prebiotics are non-digestible food ingredients that selectively stimulate the growth and activity of beneficial gut bacteria. Common prebiotics include inulin, fructooligosaccharides (FOS), and galactooligosaccharides (GOS). Prebiotics promote the development of beneficial bacteria, such as Bifidobacterium and Lactobacillus, enhancing microbial diversity and balance (64).
- Enhancing Short-Chain Fatty Acid (SCFA) Production: Prebiotics are fermented by gut bacteria to produce short-chain fatty acids (SCFAs), such as butyrate, propionate, and acetate. SCFAs play a crucial role in maintaining gut health and metabolic function. Butyrate strengthens the intestinal barrier, reduces gut permeability, and exerts anti-inflammatoryeffects (65). SCFAs also influence liver metabolism by modulating gluconeogenesis, lipogenesis, and fatty acid oxidation.
- Reducing Hepatic Steatosis and Inflammation: Prebiotics have been shown to reduce hepatic steatosis and
 inflammation in NAFLD. By improving gut microbiota composition and SCFA production, prebiotics decrease
 the influx of pro-inflammatory microbial products and modulate immune responses, reducing hepatic lipid
 accumulation and inflammation (66).
- Clinical Evidence: Clinical trials on prebiotics in NAFLD patients have shown promising results. For instance, supplementation with inulin and FOS has been associated withreductions in liver enzymes, hepatic fat content, and inflammatory markers (67). Additionally, prebiotic treatment has been linked to improved insulin sensitivity and lipid profiles in NAFLDpatients.

4.3 Combined Probiotics and Prebiotics (Synbiotics)

- Synergistic Effects: Combining probiotics and prebiotics, known as synbiotics, may offer synergistic benefits in NAFLD treatment. Synbiotics can enhance the survival and colonization of probiotics in the gut while promoting the growth of beneficial bacteria. This combined approach may more effectively modify gut microbiota, improve intestinal barrier function, and reduce hepatic inflammation (68).
- Clinical Evidence: Studies on synbiotics in NAFLD are limited but show potential benefits. For example, a synbiotic combination of Lactobacillus, Bifidobacterium, and inulin improved liver function, reduced hepatic steatosis, and decreased inflammatory markers in NAFLD patients





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Table 1: Effects of Probioticson NAFLD

Probiotic Strain	Study Type	Key Findings	Reference
Lactobacillus rhamnosus GG	ClinicalTrial	Reduced liver enzymes (ALT, AST), improved hepatic	[1]
		steatosis	
Bifidobacteriumlongum	ClinicalTrial	Decreased inflammatory markers (TNF-α, IL-	[2]
		6), improved insulin sensitivity	
Lactobacillusacidophilus	AnimalStudy	Enhanced gutbarrier integrity, reduced gut	[3]
		Permeability	
VSL#3 (multi-strain)	ClinicalTrial	al Improved liver function, reduced hepatic inflammation	

Table 2: Effects of Prebioticson NAFLD

Prebiotic	StudyType	Key Findings	Reference
Inulin	ClinicalTrial	Reduced liverenzymes, decreased hepatic fat content	[5]
Fructooligosaccharides(FOS)	ClinicalTrial	Improved insulin sensitivity, reduced inflammatory	[6]
		markers	
Galactooligosaccharides (GOS)	Animal Study	Increased SCFA production, enhanced gut barrier	[7]
		function	
Beta-glucan	Clinical Trial	Decreased liver fibrosis markers, improved lipid	[8]
		profiles	

Table 3: Effects of Synbioticson NAFLD

Synbiotic Combination	StudyType	Key Findings	Reference		
Lactobacillus +Inulin	ClinicalTrial	Reduced hepatic steatosis,improved liver function	[9]		
Bifidobacterium +FOS	ClinicalTrial	Decreased inflammatory cytokines, improved metabolic parameters	[10]		
Multi-strain Probiotic +Prebiotic	AnimalStudy	Enhanced microbial diversity, reduced liver inflammation	[11]		
Lactobacillus +Beta-glucan	ClinicalTrial	Improved gut microbiota composition, reduced liver fat	[12]		

Table 4: Mechanismsof Probiotics and Prebiotics in NAFLD

Mechanism Probiotics		Prebiotics	Synbiotics
Modulation of Gut	Increase beneficial bacteria (e.g.,	Promote growth of	Synergistic effects on gut
Microbiota	Lactobacillus, Bifidobacterium)	beneficialbacteria	microbiota composition
Enhancement of Intestinal	Upregulate tight junction	IncreaseSCFA	Combined enhancement of
Barrier	proteins,reduce permeability	production (e.g., butyrate)	barrier integrity
Reduction of Hepatic	Decrease pro- inflammatory	Modulate immune	Comprehensive reduction
Inflammation	cytokines (e.g.,TNF-α, IL-6)	responses, reduce LPS	of inflammatory markers
		translocation	
Improvement of Insulin	Improve insulin signaling	Increase GLP-1 release,	Enhanced effects on insulin
Sensitivity	pathways	improve glucose	sensitivity
		metabolism	
Reduction of Hepatic	Decrease lipid accumulation inthe	Modulate lipid	Synergistic reduction of
Steatosis	liver	metabolismpathways	liver fat content





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V. GUT BARRIER ENHANCERS: EMERGING THERAPIES TARGETING INTESTINAL BARRIER INTEGRITY IN NAFLD

The integrity of the gut barrier is crucial in preventing the translocation of microbial products that can exacerbate liver inflammation and contribute to the progression of non-alcoholic fattyliver disease (NAFLD). Emerging therapies aim to enhance gut barrier function, reduce gut permeability, and prevent the influx of harmful substances into the liver (69). Here, we reviewthe latest advancements in therapies targeting the intestinal barrier, including mucosal protectants, tight junction modulators, and microbiota-derived metabolites

5.1. Mucosal Protectants

Mucosal protectants are agents designed to shield the gut mucosa from damage, thereby preserving barrier integrity. These agents can form protective layers over the mucosal surface, reduce inflammation, and promote healing of the epithelial lining (70).

- Zinc carnosine: Zinc carnosine stabilizes the gut mucosa and has shown efficacy in enhancing mucosal defense mechanisms. It can reduce oxidative stress and inflammation, promoting the repair of the gut lining.
- Rebamipide: Originally used for treating gastric ulcers, rebamipide increases the production of mucus and prostaglandins in the gut, which can protect the epithelial barrier and enhance its healing.
- Sucralfate: This agent adheres to the ulcerated or eroded mucosal surface, creating a physical barrier that protects the gut lining from further damage and allows for repair.

5.2. Tight Junction Modulators

Tight junctions are crucial components of the intestinal barrier, regulating permeability between epithelial cells. Modulating these junctions can strengthen barrier integrity and reduce permeability (71).

- Larazotide acetate: This peptide is a tight junction regulator that prevents the disassembly of tight junction proteins, thereby reducing gut permeability. It has shown promise in clinical trials for conditions like celiac disease and could benefit NAFLD.
- Berberine: An isoquinoline alkaloid, berberine has been shown to enhance the expression of tight junction proteins such as occludin and zonula occludens-1 (ZO-1), reducing gut permeability and inflammation.
- Probiotics: Certain probiotic strains, like Lactobacillus and Bifidobacterium, can upregulate tight junction proteins, strengthening the gut barrier and reducing endotoxin translocation.

5.3. Microbiota-Derived Metabolites

Gut microbiota-derived metabolites, particularly short-chain fatty acids (SCFAs), play a significant role in maintaining gut barrier integrity (72). Enhancing the production of these beneficial metabolites can improve barrier function and reduce inflammation.

- Butyrate: A key SCFA produced by microbial fermentation of dietary fibers, butyrate has been shown to strengthen tight junctions, reduce inflammation, and promote epithelial cell health. It can be administered directly or through dietary strategies that enhance its endogenous production.
- Propionate and Acetate: These SCFAs also contribute to gut health by modulating immune responses and enhancing tight junction integrity. Increasing their levels through diet or supplements can protect the gut barrier.
- Indole derivatives: Metabolites derived from tryptophan metabolism by gut bacteria, such as indole-3-propionic acid (IPA), have been shown to enhance tight junction integrity and reduce gut permeability (73).

5.4. Prebiotic Fibers

Prebiotics are non-digestible fibers that promote the growth of beneficial gut bacteria, enhancing the production of SCFAs and other metabolites that support gut health (74).

• Inulin and Fructooligosaccharides (FOS): These prebiotics enhance the production of SCFAs, particularly butyrate, supporting gut barrier integrity and reducing inflammation.

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• Galactooligosaccharides (GOS): GOS can selectively stimulate the growth of beneficial bacteria like Bifidobacteria, leading to increased SCFA production and improved barrier function.

5.5. Phytochemicals

Phytochemicals are plant-derived compounds that can modulate gut health and barrier integrity through their antiinflammatory and antioxidant properties (75).

- Curcumin: The active compound in turmeric, curcumin, has been shown to upregulate tight junction proteins
 and reduce gut permeability. Its anti-inflammatory properties also contribute to maintaining a healthy gut
 barrier.
- Resveratrol: In grapes and berries, resveratrol can enhance tight junction integrity and reduce oxidative stress and inflammation, thereby supporting gut barrier function.

VI. CONCLUSION AND FUTURE DIRECTIONS

6.1 Conclusion:

The gut-liver axis plays a pivotal role in the pathogenesis and progression of non-alcoholic fatty liver disease (NAFLD), providing a wealth of novel insights into the mechanisms driving this complex disease. The intricate interactions between the gut microbiota, intestinal barrier integrity, bile acid metabolism, and immune responses are vital factors in developing and exacerbating NAFLD. Disruptions in the gut microbiota, commonly called dysbiosis, contribute to increased gut permeability, translocation of microbial products into the portal circulation, and subsequent hepatic inflammation. Additionally, bile acid metabolism and signaling pathway alterations further exacerbate hepatic lipid accumulation and inflammatory responses.

The therapeutic potential of targeting the gut-liver axis in NAFLD is substantial. Probiotics and prebiotics have shown promise in modulating gut microbial composition, enhancing intestinal barrier function, and reducing hepatic inflammation. Bile acid modulators, which can restore balanced bile acid signalling, also present a promising avenue for therapeutic intervention. Furthermore, gut barrier enhancers, including mucosal protectants, tight junction modulators, and microbiota-derived metabolites, can significantly reduce gut permeability and prevent the translocation of harmful substances to the liver.

By focusing on these therapeutic strategies, we can develop more effective treatments for NAFLD that address its multifaceted pathogenesis. These interventions not only have the potential to ameliorate hepatic steatosis and inflammation but also to prevent the progression to more severe forms of liver disease, such as fibrosis and cirrhosis.

6.2. Future Directions:

Future research should prioritize the following areas to further our understanding and treatment of NAFLD:

6.2.1. Elucidating Molecular Mechanisms:

Detailed investigations into the molecular mechanisms underlying the gut-liver axis are essential. This
includes studying the specific pathways through which gut microbiota influence liver metabolism and immune
responses and how alterations in these pathways contribute to NAFLD progression.

6.2.2. Identifying Key Microbial Players:

 Research should identify specific microbial species or consortia that play critical roles in promoting or mitigating NAFLD. Understanding these microbial players can lead to the development of targeted probiotic therapies.

6.2.3. Clinical Trials of Gut-Liver Axis Therapies:

Large-scale, well-designed clinical trials are needed to validate the efficacy and safety of therapeutic strategies
targeting the gut-liver axis. These trials should include diverse populations to ensure the broad applicability of
the findings.

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6.2.4. Personalized Medicine Approaches:

Given individuals' variability in gut microbiota composition, personalized medicine approaches should be explored. Tailoring therapies based on individual microbial profiles and specific pathophysiological features of NAFLD could enhance treatment outcomes.

6.2.5. Long-term Impact Studies:

Long-term studies are necessary to assess gut-liver axis interventions' sustainability and long-term impact. This includes monitoring for potential adverse effects and the durability of therapeutic benefits.

6.2.6. Combination Therapies:

Exploring combination therapies that target multiple aspects of the gut-liver axis simultaneously could provide synergistic effects and improve treatment efficacy. For instance, combining probiotics with bile acid modulators and gut barrier enhancers might yield superior results to monotherapies.

6.3. Exploring Novel Agents:

Continued exploration and development of novel agents that can modulate the gut microbiota, enhance intestinal barrier function, or modulate bile acid signaling are critical. This includes both naturally derived compounds and synthetically engineered molecules.

We can make significant strides in managing NAFLD by advancing our understanding of the gut-liver axis and translating these insights into innovative therapies. These efforts will ultimately lead to improved patient outcomes and a reduction in the burden of liver disease worldwide.

REFERENCES

- [1]. Li, J., Zou, B., Yeo, Y., Feng, Y., Xie, X., Lee, D., Fujii, H., Wu, Y., Kam, L., Ji, F., Li, X., Chien, N., Wei, M., Ogawa, E., Zhao, C., Wu, X., Stave, C., Henry, L., Barnett, S., Takahashi, H., Furusyo, N., Eguchi, Y., Hsu, Y., Lee, T., Ren, W., Qin, C., Jun, D., Toyoda, H., Wong, V., Cheung, R., Zhu, Q., & Nguyen, M. (2019). Prevalence, incidence, and outcome of non-alcoholic fatty liver disease in Asia, 1999-2019: a systematic review and meta-analysis.. The lancet. Gastroenterology & hepatology, 4 5, 389-398 . https://doi.org/10.1016/S2468-1253(19)30039-1.
- [2]. Munck, T., Xu, P., Verwijs, H., Masclee, A., Jonkers, D., Verbeek, J., & Koek, G. (2020). Intestinal permeability in human nonalcoholic fatty liver disease: A systematic review and meta □ analysis. Liver International, 40, 2906 - 2916. https://doi.org/10.1111/liv.14696.
- [3]. Michail, S., Lin, M., Frey, M., Fanter, R., Paliy, O., Hilbush, B., & Reo, N. (2015). Altered gut microbial energy and metabolism in children with non-alcoholic fatty liver disease. FEMS microbiology ecology, 91 2, 1-9 . https://doi.org/10.1093/femsec/fiu002.
- [4]. Jiang, W., Wu, N., Wang, X., Chi, Y., Zhang, Y., Qiu, X., Hu, Y., Li, J., & Liu, Y. (2015). Dysbiosis gut microbiota associated with inflammation and impaired mucosal immune function in intestine of humans with non-alcoholic fatty liver disease. Reports, 5. https://doi.org/10.1038/srep08096.
- [5]. Fianchi, F., Liguori, A., Gasbarrini, A., Grieco, A., & Miele, L. (2021). Nonalcoholic Fatty Liver Disease (NAFLD) as Model of Gut-Liver Axis Interaction: From Pathophysiology to Potential Target Treatment for Personalized Therapy. International Journal of Molecular Sciences, 22. https://doi.org/10.3390/ijms22126485.
- [6]. Carpi, R., Barbalho, S., Sloan, K., LAURINDO, L., Gonzaga, H., Grippa, P., Zutin, T., Girio, R., Repetti, C., Detregiachi, C., Bueno, P., Pereira, E., Goulart, R., & Haber, J. (2022). The Effects of Probiotics, Prebiotics and Synbiotics in Non-Alcoholic Fat Liver Disease (NAFLD) and Non-Alcoholic Steatohepatitis (NASH): A Systematic Review. International Journal of Molecular Sciences, 23. https://doi.org/10.3390/ijms23158805.





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Volume 4, Issue 1, June 2024

- [7]. Svegliati-Baroni, G., Patrício, B., Lioci, G., Macedo, M., & Gastaldelli, A. (2020). Gut- Pancreas-Liver Axis as a Target for Treatment of NAFLD/NASH. Journal of Molecular Sciences, 21. https://doi.org/10.3390/ijms21165820.
- [8]. Moore, M., Cunningham, R., Dashek, R., Mucinski, J., & Rector, R. (2020). A Fad too Far? Dietary Strategies for the Prevention and Treatment of NAFLD. Obesity, 28. https://doi.org/10.1002/oby.22964.
- [9]. Cui, Y., Wang, Q., Chang, R., Zhou, X., & Xu, C. (2019). Intestinal Barrier Function- Non-alcoholic Fatty Liver Disease Interactions and Possible Role of Gut Microbiota.. Journal of agricultural and food chemistry, 67 10, 2754-2762. https://doi.org/10.1021/acs.jafc.9b00080.
- [10]. Cui, Y., Wang, Q., Chang, R., Zhou, X., & Xu, C. (2019). Intestinal Barrier Function- Non-alcoholic Fatty Liver Disease Interactions and Possible Role of Gut Microbiota.. Journal of agricultural and food chemistry, 67 10, 2754-2762. https://doi.org/10.1021/acs.jafc.9b00080.
- [11]. Tommaso, N., Gasbarrini, A., & Ponziani, F. (2021). Intestinal Barrier in Human Health and Disease. International Journal of Environmental Research and Public Health, 18. https://doi.org/10.3390/ijerph182312836.
- [12]. Ulluwishewa, D., Anderson, R., McNabb, W., Moughan, P., Wells, J., & Roy, N. (2011). Regulation of tight junction permeability by intestinal bacteria and dietary components.. The Journal of nutrition, 141 5, 769-76. https://doi.org/10.3945/jn.110.135657.
- [13]. Kuo, W., Odenwald, M., Turner, J., & Zuo, L. (2022). Tight junction proteins occludin and ZO□1 as regulators of epithelial proliferation and survival. Academy of Sciences, 1514, 21 33. https://doi.org/10.1111/nyas.14798.
- [14]. Chen, J., & Vitetta, L. (2020). Gut Microbiota Metabolites in NAFLD Pathogenesis and Therapeutic Implications. International Journal of Molecular Sciences, 21. https://doi.org/10.3390/ijms21155214.
- [15]. Aron□Wisnewsky, J., Vigliotti, C., Witjes, J., Le, P., Holleboom, A., Verheij, J., Nieuwdorp, M., & Clément, K. (2020). Gut microbiota and human NAFLD: disentangling microbial signatures from metabolic disorders. Gastroenterology & Hepatology, 17, 279-297. https://doi.org/10.1038/s41575-020-0269-9.
- [16]. Boursier, J., Mueller, O., Barret, M., Machado, M., Fizanne, L., Araújo-Pérez, F., Guy, C., Seed, P., Rawls, J., David, L., Hunault, G., Oberti, F., Calès, P., & Diehl, A. (2016). The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. Hepatology, 63. https://doi.org/10.1002/hep.28356.
- [17]. Dokladny, K., Zuhl, M., & Moseley, P. (2016). Intestinal epithelial barrier function and tight junction proteins with heat and exercise. Journal of applied physiology, 120 6, 692-701 . https://doi.org/10.1152/japplphysiol.00536.2015.
- [18]. Miele, L., Valenza, V., Torre, G., Montalto, M., Cammarota, G., Ricci, R., Mascianà, R., Forgione, A., Gabrieli, M., Perotti, G., Vecchio, F., Rapaccini, G., Gasbarrini, G., Day, C., & Grieco, A. (2009). Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. Hepatology, 49. https://doi.org/10.1002/hep.22848.
- [19]. Tulkens, J., Vergauwen, G., Deun, J., Geeurickx, E., Dhondt, B., Lippens, L., Scheerder, M., Miinalainen, I., Rappu, P., Geest, B., Vandecasteele, K., Laukens, D., Vandekerckhove, L., Denys, H., Vandesompele, J., Wever, O., & Hendrix, A. (2018). Increased levels of systemic LPS-positive bacterial extracellular vesicles in patients with intestinal barrier dysfunction. Gut, 69, 191 193. https://doi.org/10.1136/gutjnl-2018-317726.
- [20]. Meroni, M., Longo, M., Lombardi, R., Paolini, E., Macchi, C., Corsini, A., Sirtori, C., Fracanzani, A., Ruscica, M., & Dongiovanni, P. (2021). Low Lipoprotein(a) Levels Predict Hepatic Fibrosis in Patients With Nonalcoholic Fatty Liver Disease. Hepatology Communications, 6, 535 549. https://doi.org/10.1002/hep4.1830.
- [21]. Kuo, W., Zuo, L., Odenwald, M., Madha, S., Singh, G., Gurniak, C., Abraham, C., & Turner, J. (2021). The tight junction protein ZO-1 is dispensable for barrier function but critical for effective mucosal repair.. Gastroenterology. https://doi.org/10.1053/j.gastro.2021.08.047.

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Impact Factor: 7.53

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- [22]. Ni, Y., Lu, M., Xu, Y., Wang, Q., Gu, X., Li, Y., Zhuang, T., Xia, C., Zhang, T., Gou, X., & Zhou, M. (2022). The Role of Gut Microbiota-Bile Acids Axis in the Progression of Non-alcoholic Fatty Liver Disease. Frontiers in Microbiology, 13. https://doi.org/10.3389/fmicb.2022.908011.
- [23]. Pandak, W., & Kakiyama, G. (2019). The acidic pathway of bile acid synthesis: Not just an alternative pathway. Liver research, 3, 88 98. https://doi.org/10.1016/J.LIVRES.2019.05.001.
- [24]. Pandak, W., & Kakiyama, G. (2019). The acidic pathway of bile acid synthesis: Not just an alternative pathway. Liver research, 3, 88 98. https://doi.org/10.1016/J.LIVRES.2019.05.001.
- [25]. Chiang, J., & Ferrell, J. (2020). Up to date on cholesterol 7 alpha-hydroxylase (CYP7A1) in bile acid synthesis. Liver research, 4, 47 63. https://doi.org/10.1016/j.livres.2020.05.001.
- [26]. https://doi.org/10.1016/j.jconrel.2020.07.034.
- [27]. Loomba, R., Friedman, S., & Shulman, G. (2021). Mechanisms and disease consequences of nonalcoholic fatty liver disease. Cell, 184, 2537-2564. https://doi.org/10.1016/j.cell.2021.04.015.
- [28]. Chiang, J., & Ferrell, J. (2020). Bile acid receptors FXR and TGR5 signaling in fatty liver diseases and therapy. physiology. https://doi.org/10.1152/ajpgi.00223.2019.
- [29]. Stofan, M., & Guo, G. (2020). Bile Acids and FXR: Novel Targets for Liver Diseases. Frontiers in Medicine, 7. https://doi.org/10.3389/fmed.2020.00544.
- [30]. Clifford, B., Sedgeman, L., Williams, K., Morand, P., Cheng, A., Jarrett, K., Chan, A., Brearley-Sholto, M., Wahlström, A., Ashby, J., Barshop, W., Wohlschlegel, J., Calkin, A., Liu, Y., Thorell, A., Meikle, P., Drew, B., Mack, J., Marschall, H., Tarling, E., Edwards, P., & Vallim, T. (2021). FXR activation protects against NAFLD via bile-acid-dependent reductions in lipid absorption. Cell metabolism. https://doi.org/10.1016/j.cmet.2021.06.012.
- [31]. Verbeke, L., Mannaerts, I., Schierwagen, R., Govaere, O., Klein, S., Elst, I., Windmolders, P., Farré, R., Wenes, M., Mazzone, M., Nevens, F., Grunsven, L., Trebicka, J., & Laleman, W. (2016). FXR agonist obeticholic acid reduces hepatic inflammation and fibrosis in a rat model of toxic cirrhosis. Scientific Reports, 6. https://doi.org/10.1038/srep33453.
- [32]. Zheng, C., Zhou, W., Wang, T., You, P., Zhao, Y., Yang, Y., Wang, X., Luo, J., Chen, Y., Liu, M., & Chen, H. (2015). A Novel TGR5 Activator WB403 Promotes GLP-1 Secretion and Preserves Pancreatic β-Cells in Type 2 Diabetic Mice. PLoS ONE, 10. https://doi.org/10.1371/journal.pone.0134051.
- [33]. Xie, G., Wang, X., Huang, F., Zhao, A., Chen, W., Yan, J., Zhang, Y., Lei, S., Ge, K., Zheng, X., Liu, J., Su, M., Liu, P., & Jia, W. (2016). Dysregulated hepatic bile acids collaboratively promote liver carcinogenesis. International Journal of Cancer, 139. https://doi.org/10.1002/ijc.30219.
- [34]. Linden, A., Li, S., Choi, H., Fang, F., Fukasawa, M., Uyeda, K., Hammer, R., Horton, J., Engelking, L., & Liang, G. (2018). Interplay between ChREBP and SREBP-1c coordinates postprandial glycolysis and lipogenesis in livers of mice[S]. Lipid Research, 59, 475 487. https://doi.org/10.1194/jlr.M081836.
- [35]. Nguyen, T., Kim, D., Lee, Y., Lee, Y., Truong, X., Lee, J., Song, D., Kwon, T., Park, S., Jung, C., Moon, C., Osborne, T., Im, S., & Jeon, T. (2021). SREBP-1c impairs ULK1 sulfhydration-mediated autophagic flux to promote hepatic steatosis in high-fat-diet- fed mice.. Molecular cell. https://doi.org/10.1016/j.molcel.2021.06.003.
- [36]. Tanaka, M., Kishimoto, Y., Sasaki, M., Sato, A., Kamiya, T., Kondo, K., & Iida, K. (2018). Terminalia bellirica (Gaertn.) Roxb. Extract and Gallic Acid Attenuate LPS- Induced Inflammation and Oxidative Stress via MAPK/NF-κB and Akt/AMPK/Nrf2 Pathways. Oxidative Medicine and Cellular Longevity, 2018. https://doi.org/10.1155/2018/9364364.
- [37]. Chávez-Talavera, O., Tailleux, A., Lefebvre, P., & Staels, B. (2017). Bile acid control of metabolism and inflammation in obesity, type 2 diabetes, dyslipidemia and NAFLD. Gastroenterology.
- [38]. Jiang, W., Wu, N., Wang, X., Chi, Y., Zhang, Y., Qiu, X., Hu, Y., Li, J., & Liu, Y. (2015). Dysbiosis gut microbiota associated with inflammation and impaired mucosal immune function in intestine of humans with non-alcoholic fatty liver disease. Reports, 5. https://doi.org/10.1038/srep08096.





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Impact Factor: 7.53

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- [39]. Amer, J., Salhab, A., Noureddin, M., Doron, S., Abu-Tair, L., Ghantous, R., Mahamid, M., & Safadi, R. (2018). Insulin signaling as a potential natural killer cell checkpoint in fatty liver disease. Hepatology Communications, 2, 285 298. https://doi.org/10.1002/hep4.1146.
- [40]. Duan, Y., Pan, X., Luo, J., Xiao, X., Li, J., Bestman, P., & Luo, M. (2022). Association of Inflammatory Cytokines With Non-Alcoholic Fatty Liver Disease. Immunology, 13. https://doi.org/10.3389/fimmu.2022.880298.
- [41]. Chen, J., Deng, X., Liu, Y., Tan, Q., Huang, G., Che, Q., Guo, J., & Su, Z. (2020). Kupffer Cells in Non-alcoholic Fatty Liver Disease: Friend or Foe?. Journal of Biological Sciences, 16, 2367 2378. https://doi.org/10.7150/ijbs.47143.
- [42]. Slevin, E., Baiocchi, L., Wu, N., Ekser, B., Sato, K., Lin, E., Ceci, L., Chen, L., Lorenzo, S., Xu, W., Kyritsi, K., Meadows, V., Zhou, T., Kundu, D., Han, Y., Kennedy, L., Glaser, S., Francis, H., Alpini, G., & Meng, F. (2020). Kupffer cells: Inflammation pathways and cell-cell interactions in alcohol-associated liver disease.. pathology. https://doi.org/10.1016/j.ajpath.2020.08.014.
- [43]. Alabdulaali, B., Al-Rashed, F., Al-Onaizi, M., Kandari, A., Razafiarison, J., Tonui, D., Williams, M., Blériot, C., Ahmad, R., & Alzaid, F. (2023). Macrophages and the development and progression of non-alcoholic fatty liver disease. Immunology, 14. https://doi.org/10.3389/fimmu.2023.1195699.
- [44]. Tacke, F., & Zimmermann, H. (2014). Macrophage heterogeneity in liver injury and fibrosis. Journal of hepatology, 60 5, 1090-6. https://doi.org/10.1016/j.jhep.2013.12.025.
- [45]. Kaltenmeier, C., Yazdani, H., Handu, S., Popp, B., Geller, D., & Tohme, S. (2022). The Role of Neutrophils as a Driver in Hepatic Ischemia-Reperfusion Injury and Cancer Growth. Frontiers in Immunology, 13. https://doi.org/10.3389/fimmu.2022.887565.
- [46]. Her, Z., Tan, J., Lim, Y., Tan, S., Chan, X., Tan, W., Liu, M., Yong, K., Lai, F., Ceccarello, E., Zheng, Z., Fan, Y., Chang, K., Sun, L., Chang, S., Chin, C., Lee, G., Dan, Y., Chan, Y., Lim, S., Chan, J., Chandy, K., & Chen, Q. (2020). CD4+ T Cells Mediate the Development of Liver Fibrosis in High Fat Diet-Induced NAFLD in Humanized Mice. Frontiers in Immunology, 11. https://doi.org/10.3389/fimmu.2020.580968.
- [47]. Perugino, C., Kaneko, N., Maehara, T., Mattoo, H., Kers, J., Allard-Chamard, H., Mahajan, V., Liu, H., Della-Torre, E., Murphy, S., Ghebremichael, M., Wallace, Z., Bolster, M., Harvey, L., Mylvaganam, G., Tuncay, Y., Liang, L., Montesi, S., Zhang, X., Tinju, A., Mochizuki, K., Munemura, R., Sakamoto, M., Moriyama, M., Nakamura, S., Yosef, N., Stone, J., & Pillai, S. (2020). CD4+ and CD8+ cytotoxic T lymphocytes may induce mesenchymal cell apoptosis in IgG4-related disease.. allergy and clinical immunology. https://doi.org/10.1016/j.jaci.2020.05.022.
- [48]. Diedrich, T., Kummer, S., Galante, A., Drolz, A., Schlicker, V., Lohse, A., Kluwe, J., Eberhard, J., & Wiesch, J. (2020). Characterization of the immune cell landscape of patients with NAFLD. PLoS ONE, 15. https://doi.org/10.1371/journal.pone.0230307.
- [49]. Wang, Z., Liang, X., Xiong, A., Ding, L., Li, W., Yang, L., Wu, X., Shi, H., Zhou, Y., & Wang, Z. (2021). Helichrysetin and TNF-α synergistically promote apoptosis by inhibiting overactivation of the NF-κB and EGFR signaling pathways in HeLa and T98G cells. International Journal of Molecular Medicine, 47. https://doi.org/10.3892/ijmm.2021.4882.
- [50]. Ahmed, Y., Fu, Y., Rodrigues, R., He, Y., Guan, Y., Guillot, A., Ren, R., Feng, D., Hidalgo, J., Ju, C., Lafdil, F., & Gao, B. (2021). Kupffer cell restoration after partial hepatectomy is mainly driven by local cell proliferation in IL-6-dependent autocrine and paracrine manners. Cellular & Molecular Immunology, 18, 2165 2176. https://doi.org/10.1038/s41423-021-00731-7.
- [51]. Johnson, D., O'Keefe, R., & Grandis, J. (2018). Targeting the IL-6/JAK/STAT3 signalling axis in cancer. Nature Reviews Clinical Oncology, 15, 234-248. https://doi.org/10.1038/nrclinonc.2018.8.
- [52]. Robert, S., Gicquel, T., Bodin, A., Lagente, V., & Boichot, E. (2016). Characterization of the MMP/TIMP Imbalance and Collagen Production Induced by IL-1β or TNF-α Release from Human Hepatic Stellate Cells. PLoS ONE, 11. https://doi.org/10.1371/journal.pone.0153118.





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International Open-Access, Double-Blind, Peer-Reviewed, Refereed, Multidisciplinary Online Journal

Impact Factor: 7.53

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- [53]. Baeck, C., Wehr, A., Karlmark, K., Heymann, F., Vucur, M., Gassler, N., Huss, S., Klußmann, S., Eulberg, D., Luedde, T., Trautwein, C., & Tacke, F. (2011). Pharmacological inhibition of the chemokine CCL2 (MCP-1) diminishes liver macrophage infiltration and steatohepatitis in chronic hepatic injury. Gut, 61, 416 -426. https://doi.org/10.1136/gutjnl-2011-300304.
- [54]. Barnabei, L., Laplantine, E., Mbongo, W., Rieux-Laucat, F., & Weil, R. (2021). NF-κB: At the Borders of Autoimmunity and Inflammation. Frontiers in Immunology, 12. https://doi.org/10.3389/fimmu.2021.716469.
- [55]. Xin, P., Xu, X., Deng, C., Liu, S., Wang, Y., Zhou, X., Ma, H., Wei, D., & Sun, S. (2020). The role of JAK/STAT signaling pathway and its inhibitors in diseases.. International immunopharmacology, 80, 106210 . https://doi.org/10.1016/j.intimp.2020.106210.
- [56]. Wree, A., McGeough, M., Inzaugarat, M., Eguchi, A., Schuster, S., Johnson, C., Peña, C., Geisler, L., Papouchado, B., Hoffman, H., & Feldstein, A. (2018). NLRP3 inflammasome driven liver injury and fibrosis: Roles of IL□17 and TNF in mice. Hepatology, 67. https://doi.org/10.1002/hep.29523.
- [57]. Ji, Y., Yin, Y., Sun, L., & Zhang, W. (2020). The Molecular and Mechanistic Insights Based on Gut-Liver Axis: Nutritional Target for Non-Alcoholic Fatty Liver Disease (NAFLD) Improvement. International Journal of Molecular Sciences, 21. https://doi.org/10.3390/ijms21093066.
- [58]. Marzorati, M., Abbeele, P., Bubeck, S., Bayne, T., Krishnan, K., & Young, A. (2021). Treatment with a spore-based probiotic containing five strains of Bacillus induced changes in the metabolic activity and community composition of the gut microbiota in a SHIME® model of the human gastrointestinal system.. Food research international, 149, 110676. https://doi.org/10.1016/j.foodres.2021.110676.
- [59]. Liang, Y., Liang, S., Zhang, Y., Deng, Y., He, Y., Chen, Y., Liu, C., Lin, C., & Yang, O. (2018). Oral Administration of Compound Probiotics Ameliorates HFD-Induced Gut Microbe Dysbiosis and Chronic Metabolic Inflammation via the G Protein-Coupled Receptor 43 in Non-alcoholic Fatty Liver Disease Rats. Proteins, 11, 175-185. https://doi.org/10.1007/s12602-017-9378-3.
- [60]. Wu, Y., Jha, R., Li, A., Liu, H., Zhang, Z., Zhang, C., Zhai, Q., & Zhang, J. (2022). Probiotics (Lactobacillus plantarum HNU082) Supplementation Relieves Ulcerative Colitis by Affecting Intestinal Barrier Functions, Immunity-Related Gene Expression, Gut Microbiota, and Metabolic Pathways in Mice. Microbiology Spectrum, 10. https://doi.org/10.1128/spectrum.01651-22.
- [61]. Munck, T., Xu, P., Verwijs, H., Masclee, A., Jonkers, D., Verbeek, J., & Koek, G. (2020). Intestinal permeability in human nonalcoholic fatty liver disease: A systematic review and meta □ analysis. Liver International, 40, 2906 - 2916. https://doi.org/10.1111/liv.14696.
- [62]. Liu, Q., Tian, H., Kang, Y., Tian, Y., Li, L., Kang, X., Yang, H., Wang, Y., Tian, J., Zhang, F., Tong, M., Cai, H., & Fan, W. (2021). Probiotics alleviate autoimmune hepatitis in mice through modulation of gut microbiota and intestinal permeability.. 108863. https://doi.org/10.1016/j.jnutbio.2021.108863.
- [63]. Pinto-Sanchez, M., Hall, G., Ghajar, K., Nardelli, A., Bolino, C., Lau, J., Martin, F., Cominetti, O., Welsh, C., Rieder, A., Traynor, J., Gregory, C., Palma, G., Pigrau, M., Ford, A., Macri, J., Berger, B., Bergonzelli, G., Surette, M., Collins, S., Moayyedi, P., & Bercik, P. (2017). Probiotic Bifidobacterium longum NCC3001 Reduces Depression Scores and Alters Brain Activity: A Pilot Study in Patients With Irritable Bowel Syndrome.. Gastroenterology, 153 2, 448-459.e8 . https://doi.org/10.1053/j.gastro.2017.05.003.
- [64]. Carlson, J., Erickson, J., Lloyd, B., & Slavin, J. (2018). Health Effects and Sources of Prebiotic Dietary Fiber. Current Developments in Nutrition, 2. https://doi.org/10.1093/cdn/nzy005.
- [65]. Yang, Z., Guo, Y., Huang, J., Gao, Y., Peng, F., Xu, R., Su, H., & Zhang, P. (2021). Isomaltulose Exhibits Prebiotic Activity, and Modulates Gut Microbiota, the Production of Short Chain Fatty Acids, and Secondary Bile Acids in Rats. Molecules, 26. https://doi.org/10.3390/molecules26092464.
- [66]. Zhong, M., Yan, Y., Yuan, H., A, R., Xu, G., Cai, F., Yang, Y., Wang, Y., & Zhang, W. (2022). Astragalus mongholicus polysaccharides ameliorate hepatic lipid accumulation and inflammation as well as modulate gut microbiota in NAFLD rats.. Food & function. https://doi.org/10.1039/d2fo01009g.
- [67]. Carpi, R., Barbalho, S., Sloan, K., LAURINDO, L., Gonzaga, H., Grippa, P., Zutin, T., Girio, R., Repetti, C., Detregiachi, C., Bueno, P., Pereira, E., Goulart, R., & Haber, J. (2022). The Effects of Robiotics, Prebiotics

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- and Synbiotics in Non-Alcoholic Fat Liver Disease (NAFLD) and Non-Alcoholic Steatohepatitis (NASH): A Systematic Review. International Journal of Molecular Sciences, 23. https://doi.org/10.3390/ijms23158805.
- [68]. Scorletti, E., Afolabi, P., Miles, E., Smith, D., Almehmadi, A., Alshathry, A., Childs, C., Fabbro, S., Bilson, J., Moyses, H., Clough, G., Sethi, J., Patel, J., Wright, M., Breen, D., Peebles, C., Darekar, A., Aspinall, R., Fowell, A., Dowman, J., Nobili, V., Targher, G., Delzenne, N., Bindels, L., Calder, P., & Byrne, C. (2020). Synbiotic Alters Fecal Microbiomes, but not Liver Fat or Fibrosis, in a Randomized Trial of Patients With Non-alcoholic Fatty Liver Disease.. Gastroenterology. https://doi.org/10.1053/j.gastro.2020.01.031.
- [69]. Craven, L., Rahman, A., Parvathy, S., Beaton, M., Silverman, J., Qumosani, K., Hramiak, I., Hegele, R., Joy, T., Meddings, J., Urquhart, B., Harvie, R., McKenzie, C., Summers, K., Reid, G., Burton, J., & Silverman, M. (2020). Allogenic Fecal Microbiota Transplantation in Patients With Nonalcoholic Fatty Liver Disease Improves Abnormal Small Intestinal Permeability: A Randomized Control Trial.. gastroenterology. https://doi.org/10.14309/ajg.0000000000000661.
- [70]. Saia, R., Ribeiro, A., & Giusti, H. (2020). Cholecystokinin Modulates the Mucosal Inflammatory Response and Prevents the Lipopolysaccharide-Induced Intestinal Epithelial Barrier Dysfunction.. Shock. https://doi.org/10.1097/SHK.0000000000001355.
- [71]. Paradis, T., Bègue, H., Basmaciyan, L., Dalle, F., & Bon, F. (2021). Tight Junctions as a Key for Pathogens Invasion in Intestinal Epithelial Cells. Molecular Sciences, 22. https://doi.org/10.3390/ijms22052506.
- [72]. Gasaly, N., Vos, P., & Hermoso, M. (2021). Impact of Bacterial Metabolites on Gut Barrier Function and Host Immunity: A Focus on Bacterial Metabolism and Its Relevance for Intestinal Inflammation. Frontiers in Immunology, 12. https://doi.org/10.3389/fimmu.2021.658354.
- [73]. Jiang, H., Chen, C., & Gao, J. (2022). Extensive Summary of the Important Roles of Indole Propionic Acid, a Gut Microbial Metabolite in Host Health and Disease. Nutrients, 15. https://doi.org/10.3390/nu15010151.
- [74]. Venegas, D., Fuente, M., Landskron, G., González, M., Quera, R., Dijkstra, G., Harmsen, H., Faber, K., & Hermoso, M. (2019). Short Chain Fatty Acids (SCFAs)- Mediated Gut Epithelial and Immune Regulation and Its Relevance for Inflammatory Bowel Diseases. Frontiers in Immunology, 10. https://doi.org/10.3389/fimmu.2019.00277.
- [75]. Ruan, D., Wu, S., Fouad, A., Zhu, Y., Huang, W., Chen, Z., Gou, Z., Wang, Y., Han, Y., Yan, S., Zheng, C., & Jiang, S. (2022). Curcumin alleviates LPS-induced intestinal homeostatic imbalance through reshaping gut microbiota structure and regulating group3 innate lymphoid cells in chickens. Food & function. https://doi.org/10.1039/d2fo02598a.

REFERENCES FOR TABLES

- [1]. Famouri, F., Shariat, Z., Hashemipour, M., Keikha, M., & Kelishadi, R. (2017). Effects of Probiotics on Nonalcoholic Fatty Liver Disease in Obese Children and Adolescents. Journal of Pediatric Gastroenterology and Nutrition, 64, 413–417. https://doi.org/10.1097/MPG.00000000001422.
- [2]. Malaguarnera, M., Vacante, M., Antić, T., Giordano, M., Chisari, G., Acquaviva, R., Mastrojeni, S., Malaguarnera, G., Mistretta, A., Volti, G., & Galvano, F. (2012). Bifidobacterium longum with Fructo-Oligosaccharides in Patients with Non Alcoholic Steatohepatitis. Digestive Diseases and Sciences, 57, 545-553.
 - https://doi.org/10.1007/s10620-011-1887-4.
- [3]. Koutnikova, H., Genser, B., Monteiro-Sepulveda, M., Faurie, J., Rizkalla, S., Schrezenmeir, J., & Clément, K. (2019). Impact of bacterial probiotics on obesity, diabetes and non-alcoholic fatty liver disease related variables: a systematic review and meta-analysis of randomised controlled trials. BMJ Open, 9. https://doi.org/10.1136/bmjopen-2017-017995.
- [4]. Koutnikova, H., Genser, B., Monteiro-Sepulveda, M., Faurie, J., Rizkalla, S., Schrezenmeir, J., & Clément, K. (2019). Impact of bacterial probiotics on obesity, diabetes and non-alcoholic fatty liver disease related variables: a systematic review and meta-analysis of randomised controlled trials. BMJ Open, 9. https://doi.org/10.1136/bmjopen-2017-017995.

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- [5]. Aliasgharzadeh, A., Khalili, M., Mirtaheri, E., Gargari, B., Tavakoli, F., Farhangi, M., Babaei, H., & Dehghan, P. (2015). A Combination of Prebiotic Inulin and Oligofructose Improve Some of Cardiovascular Disease Risk Factors in Women with Type 2 Diabetes: A Randomized Controlled Clinical Trial.. Advanced pharmaceutical bulletin, 5 4, 507- 14. https://doi.org/10.15171/apb.2015.069.
- [6]. Liu, Y., Chen, K., Li, F., Gu, Z., Liu, Q., He, L., Shao, T., Song, Q., Zhu, F., Zhang, L., Jiang, M., Zhou, Y., Barve, S., Zhang, X., McClain, C., & Feng, W. (2020). Probiotic Lactobacillus rhamnosus GG Prevents Liver Fibrosis Through Inhibiting Hepatic Bile Acid Synthesis and Enhancing Bile Acid Excretion in Mice. Md.), 71, 2050 2066. https://doi.org/10.1002/hep.30975.
- [7]. Doulberis, M., Kotronis, G., Gialamprinou, D., Kountouras, J., & Katsinelos, P. (2017). Non-alcoholic fatty liver disease: An update with special focus on the role of gut microbiota.. Metabolism: clinical and experimental, 71, 182-197. https://doi.org/10.1016/j.metabol.2017.03.013.
- [8]. Baker, M., Cho, B., Anez-Bustillos, L., Dao, D., Pan, A., O'Loughlin, A., Lans, Z., Mitchell, P., Nosé, V., Gura, K., Puder, M., & Fell, G. (2019). Fish oil-based injectable lipid emulsions containing medium-chain triglycerides or added α-tocopherol offer anti-inflammatory benefits in a murine model of parenteral nutrition-induced liver injury. The American journal of clinical nutrition, 109 4, 1038-1050 . https://doi.org/10.1093/ajcn/nqy370.
- [9]. Scorletti, E., & Byrne, C. (2013). Omega-3 fatty acids, hepatic lipid metabolism, and nonalcoholic fatty liver disease.. Annual review of nutrition, 33, 231-48. https://doi.org/10.1146/annurev-nutr-071812-161230.
- [10]. Xue, L., He, J., Gao, N., Lu, X., Li, M., Wu, X., Liu, Z., Jin, Y., Liu, J., Xu, J., & Geng, Y. (2017). Probiotics may delay the progression of nonalcoholic fatty liver disease by restoring the gut microbiota structure and improving intestinal endotoxemia. Reports, 7. https://doi.org/10.1038/srep45176.
- [11]. Carpi, R., Barbalho, S., Sloan, K., LAURINDO, L., Gonzaga, H., Grippa, P., Zutin, T., Girio, R., Repetti, C., Detregiachi, C., Bueno, P., Pereira, E., Goulart, R., & Haber, J. (2022). The Effects of Probiotics, Prebiotics and Synbiotics in Non-Alcoholic Fat Liver Disease (NAFLD) and Non-Alcoholic Steatohepatitis (NASH): A Systematic Review. International Journal of Molecular Sciences, 23. https://doi.org/10.3390/ijms23158805.
- [12]. Bakhshimoghaddam, F., Shateri, K., Sina, M., Hashemian, M., & Alizadeh, M. (2018). Daily Consumption of Synbiotic Yogurt Decreases Liver Steatosis in Patients with Nonalcoholic Fatty Liver Disease: A Randomized Controlled Clinical Trial.. Journal of nutrition, 148 8, 1276-1284 . https://doi.org/10.1093/jn/nxy088.
- [13]. Fan, J., & Farrell, G. (2009). Epidemiology of non-alcoholic fatty liver disease in China.. Journal of hepatology, 50 1, 204-10 . https://doi.org/10.1016/j.jhep.2008.10.010.
- [14]. Luck, H., Tsai, S., Chung, J., Clemente-Casares, X., Ghazarian, M., Revelo, X., Lei, H., Luk, C., Shi, S., Surendra, A., Copeland, J., Ahn, J., Prescott, D., Rasmussen, B., Chng, M., Engleman, E., Girardin, S., Lam, T., Croitoru, K., Dunn, S., Philpott, D., Guttman, D., Woo, M., Winer, S., & Winer, D. (2015). Regulation of obesity-related insulin resistance with gut anti-inflammatory agents.. Cell metabolism, 21 4, 527-42 . https://doi.org/10.1016/j.cmet.2015.03.001.
- [15]. Spruss, A., & Bergheim, I. (2009). Dietary fructose and intestinal barrier: potential risk factor in the pathogenesis of nonalcoholic fatty liver disease.. biochemistry, 20 9, 657-62 . https://doi.org/10.1016/j.jnutbio.2009.05.006.
- [16]. Wallace, T., Guarner, F., Madsen, K., Cabana, M., Gibson, G., Hentges, E., & Sanders, M. (2011). Human gut microbiota and its relationship to health and disease.. reviews, 69 7, 392-403 . https://doi.org/10.1111/j.1753-4887.2011.00402.x.
- [17]. Huda, M., Salvador, A., Barrington, W., Gacasan, C., D'Souza, E., Ramirez, L., Threadgill, D., & Bennett, B. (2022). Gut microbiota and host genetics modulate the effect of diverse diet patterns on metabolic health. Frontiers in Nutrition, 9. https://doi.org/10.3389/fnut.2022.896348.

